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(54) Title: IDENTIFICATION OF THE GENETIC DETERMINANTS OF THE POLYMORPHIC CYP3A5 EXPRESSION

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(57) Abstract: The present invention relates to a polymorphic CYP3A5 polynucleotide. Moreover, the invention relates to genes or vectors comprising the polynucleotides of the invention and to a host cell genetically engineered with the polynucleotide or gene of the invention. Further, the invention relates to methods for producing molecular variant polypeptides or fragments thereof, methods for producing calls capable of expressing a molecular variant polypeptide and to a polypeptide or fragment thereof encoded by the polynucleotide or the gene of the invention or which is obtainable by the method or from the cells produced by the method of the invention. Furthermore, the invention relates to an antibody which binds specifically the polapeptide of the invention. Moreover, the invention relates to a transgenic non-human animal. The invention also relates to a solid support comprising one or a plurality of the above mentioned polynucleotides, genes, vectors, polypeptides, antibodies or host cells. Furthermore, methods of identifying a polymorphism, identifying and obtaining a prodrug or drug or an inhibitor are also encompassed by the present invention. In addition, the invention relates to methods for producing of a pharmaceutical composition and methods of diagnosing a disease. Further, the invention relates to a method of detection of the polynucleotide of the invention. Furthermore, comprised by the present invention are a diagnostic and a pharmaceutical composition. Even more the invention relates to uses of the polynucleotides, genes, vectors, polypeptides or antibodies of the invention. Finally, the invention relates to a diagnostic kit.

Identification of the genetic determinants of the polymorphic CYP3A5 expression

The present invention relates to a polymorphic CYP3A5 polynucleotide. Moreover, the invention relates to genes or vectors comprising the polynucleotides of the invention and to a host cell genetically engineered with the polynucleotide or gene of the invention. Further, the invention relates to methods for producing molecular variant polypeptides or fragments thereof, methods for producing cells capable of expressing a molecular variant polypeptide and to a polypeptide or fragment thereof encoded by the polynucleotide or the gene of the invention or which is obtainable by the method or from the cells produced by the method of the invention. Furthermore, the invention relates to an antibody which binds specifically the polypeptide of the invention. Moreover, the invention relates to a transgenic non-human animal. The invention also relates to a solid support comprising one or a plurality of the above mentioned polynucleotides, genes, vectors, polypeptides, antibodies or host cells. Furthermore, methods of identifying a polymorphism, identifying and obtaining a pro-drug or drug or an inhibitor are also encompassed by the present invention. In addition, the invention relates to methods for producing of a pharmaceutical composition and to methods of diagnosing a disease. Further, the invention relates to a method of detection of the polynucleotide of the invention. Furthermore, comprised by the present invention are a diagnostic and a pharmaceutical composition. Even more, the invention relates to uses of the polynucleotides, genes, vectors, polypeptides or antibodies of the invention. Finally, the invention relates to a diagnostic kit.

The CYP3A enzymes play a particularly important role in drug metabolism. This is due to their abundant expression in the liver combined with a broad substrate spectrum. Indeed, it is estimated that CYP3A isozymes collectively comprise the largest portion of the liver CYP protein (Thummel, Annu Rev Pharmacol Toxicol 38 (1998), 389-430) and that they are involved in the metabolism of 45 % - 60 % of all currently used drugs (Li, Toxicology 104 (1995), 1-8; Evans, Science 286 (1999), 487-91). In addition to drugs, CYP3A isozymes metabolise a variety of other compounds including steroid hormones, toxins and carcinogens. For example, CYP3A isozymes metabolise aflatoxin B₁ (Wang, Biochemistry

37 (1998), 12536 - 45; Gillam, Arch Biochem Biophys 317 (1995), 374-84; Li, Cancer Res 57 (1997), 641-5), a mycotoxin strongly implicated in the etiology of liver cancer, which is a major cause of premature death in many areas of Africa and Asia (Henry, Science 286 (1999), 2453-4).

The hepatic expression and activity of CYP3A isozymes is inter-individually variable and this variability is the reason for harmful interactions frequently encountered in development and application of drugs that are CYP3A substrates. It has also been postulated that variable CYP3A expression could affect an individual's predisposition to cancers caused by environmental carcinogens which are metabolised by CYP3A. The elucidation of factors controlling an individual's CYP3A activity could permit personalised dose adjustments in therapies with its substrates and also lead to the identification of sub-populations at increased risk for several common cancers. However, despite considerable efforts, our understanding of factors governing CYP3A activity and expression is limited. There are several reasons for this: An average human liver may express products of up to four CYP3A genes (Gellner, Pharmacogenetics 11 (2001), 111 - 121), but their respective contributions to the hepatic CYP3A pool are still a matter of debate. The differentiation between the individual CYP3A proteins by enzymatic methods has proven difficult due to overlapping substrate specifities and due to the considerable effect of reconstitution. conditions on their catalytic activities. RNA and protein analysis indicate that CYP3A4 forms the bulk of the hepatic CYP3A protein and its expression is highly variable (Thummel, Annu Rev Pharmacol Toxicol 38 (1998), 389-430). Less well understood are the contributions of the other CYP3A genes. CYP3A5 is widely considered the second most important CYP3A protein in the liver, but the available data are conflicting, since its expression has been reported to be present in 10 % to 97 % of human livers (Aoyama, J Biol Chem 264 (1989), 10388-95; Wrighton, Mol Pharmacol 38 (1990), 207-13; Schuetz, Pharmacogenetics 4 (1994), 11-20; Jounaidi, Biochem Biophys Res Commun 221 (1996). 466-70; Boobis, Br J Clin Pharmacol 42 (1996), 81-9). The possible reasons for these discrepancies include small sample sizes, interethnic differences and poor specificity of probes used to measure CYP3A5 expression. The third CYP3A, CYP3A7, was originally described in the human fetal liver where it accounts for about 50 % of the total CYP protein (Wrighton, Biochem Pharmacol 37 (1988), 3053-5). More recent studies indicate constitutive or induced expression of CYP3A7 in adult human livers, but its quantification has been hampered by the lack of specific antibodies. Similarly, no protein expression

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data are available for the recently identified fourth member of the family, CYP3A43 (Gellner, Pharmacogenetics 11 (2001), 111 - 121).

Clinical studies indicate that a major portion of the inter-individual CYP3A variability is caused by genetic factors (Ozdemir, Pharmacogenetics 10 (2000), 373-88), but the identities of the latter remain unknown. In respect of CYP3A5, a protein variant (Thr398Asn) has been found in 2 out of 5 individuals deficient in CYP3A5 expression (Jounaidi, Biochem Biophys Res Commun 221 (1996), 466-70), but its significance has not been verified on a larger number of liver samples and in functional studies. In addition, a haplotype consisting of two linked polymorphisms has been described in the 5' flanking region of the CYP3A5 gene which is associated with increased expression and activity of the gene (Paulussen, Pharmacogenetics 10 (2000), 415-24). However, only a small sample set (n=29) was analysed for the genotype and the phenotype. Moreover, the single nucleotide polymorphisms (SNPs) which have been disclosed in said document are not suitable for a reliable prediction of CYP3A5 dysfunction and/or dysregulation and the problems caused thereby. This document does not suggest the existence of further haplotypes.

Thus, improved means and methods for diagnosing and treating a variety of diseases and disorders based on dysfunctions or dysregulations of drug metabolism were not available yet but are nevertheless highly desirable. Thus, the technical problem underlying the present invention is to comply with the above specified needs.

The solution to this technical problem is achieved by providing the embodiments characterized in the claims.

Accordingly, the present invention relates to a polynucleotide comprising a polynucleotide selected from the group consisting of:

(a) a polynucleotide having the nucleic acid sequence of SEQ ID NO: 54, 56, 58, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94, 96, 98, 100, 102, 106, 108, 110, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 128, 129, 130, 131, 133, 134, 135, 136, 137, 138, 139, 140, 142, 143, 149, 151, 153, 155, 157, 159, 161, 163, 165, 169, 171, 173, 175, 177, 179, 181, 183, 185, 187, 189, 193, 195, 197, 199, 201, 207, 208,

209, 210, 211, 212, 213, 214, 216, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 231, 232, 233, 235, or 236;

- (b) a polynucleotide encoding a polypeptide having the amino acid sequence of SEQ ID NO: 127, 132, 141, 215, 229, or 234;
- (c) a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said polynucleotide is having a nucleotide exchange, a nucleotide deletion of at least one nucleotide, or at least one additional nucleotide at a position corresponding to position -20643, -20555, -20359, -20367, -20329, -20323, -20310, -6200, -6177, -4336, -3990, -3868, -3844, -3557, -1617, -795, -86, -74, 136, 174 to 176, 230, 3705, 3709/3710, 5215, 5235, 5516, 7182, 7207, 7303, 7424/7427, 12907, 13028, 13077, 13173, 13226, 13376, 14720, 14836, 14903, 15788, 16079, 16931/16932, 16993, 17163, 19069, 19165, 19208, 27050, 27131/27132, 27526, 31499, 31551 or 31611 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614);
- a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said (d) polynucleotide is having an A at a position corresponding to position -20555, -20329, -20323, -4336, -3868, -3844, -795, -86, 230, 5235, 5516, 7182, 7303, 12907, 13028, 13376, 19069 or 19165 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a T at a position corresponding to position -20367, -6200, -74, 3705, 5215, 7207, 14836, 17163, 19208 or 27526 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a G at a position corresponding to position -6177, -3990, 13077, 14720, 14903, 16993 or 27050 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a C at a position corresponding to position -20643, -20310, -3557, -1617, 136, 13173, 13226, 15788, 16079, 31499, 31551 or

31611 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), nucleotide deletions at positions corresponding to positions 174 to 176 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613), an additional nucleotide at a position corresponding to position 3709/3710 or 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), three additional nucleotides at a position corresponding to position 16931/16932 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), or a deletion of two nucleotides and nine additional nucleotides inserted at a position corresponding to position 7424 to 7427 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613);

- (e) a polynucleotide encoding a CYP3A5 polypeptide or fragment thereof, wherein said polypeptide comprises an amino acid substitution at a position corresponding to position 30, 100, 130, 149 or 488 of the CYP3A5 polypeptide (Accession No: NP_000768.1), or at least one amino acid exchange or a stop codon at a position corresponding to position 30 to 34 or 346 to 348 of the CYP3A5 polypeptide (Accession No: NP_000768.1); and
- a polynucleotide encoding a CYP3A5 polypeptide or fragment thereof, wherein said polypeptide comprises amino acid substitutions of HGLFK to YGTF (with the period meaning termination) at a position corresponding to position 30 to 34 of the CYP3A5 polypeptide (Accession No: NP_000768.1, an amino acid substitution of S to Y at a position corresponding to position 100 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid substitution of R to Q at a position corresponding to position 130 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid substitution of I to T at a position corresponding to position 149 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid

substitutions of TYD to YL. (with the period meaning termination) at position corresponding to position 346 to 348 of the CYP3A5 polypeptide (Accession No: NP_000768.1), or an amino acid substitution of I to T at a position corresponding to position 488 of the CYP3A5 polypeptide (Accession No: NP_000768.1).

In the context of the present invention the term "polynucleotides" or the term "polypeptides" refers to different variants of a polynucleotide or polypeptide. Said variants comprise a reference or wild type sequence of the polynucleotides or polypeptides of the invention as well as variants which differ therefrom in structure or composition. Reference or wild type sequences for the polynucleotides are Accession No: AF280107.1 and AC005020.2. Reference or wild type sequence for the polypeptides of the invention is Accession No: NP 000768.1. The differences in structure or composition usually occur by way of nucleotide or amino acid substitution(s), addition(s) and/or deletion(s). Preferably, said nucleotide substitution(s), addition(s) or deletion(s) result(s) in one or more changes of the corresponding amino acid(s) of the polypeptides of the invention. The variant polynucleotides and polypeptides also comprise fragments of said polynucleotides or polypeptides of the invention. The polynucleotides and polypeptides as well as the aforementioned fragments thereof of the present invention are characterized as beingassociated with a CYP3A5 dysfunction or dysregulation. Preferably, said dysfunctions or dysregulations referred to in the present invention cause a disease or disorder or a prevalence for said disease or disorder. Preferably, as will be discussed below in detail, said disease is cancer or diseases including cardiovascular diseases, diabetes and AIDS or any other disease caused by a dysfunction or dysregulation due to a polynucleotide or polypeptides of the invention.

The polynucleotides of the invention include polynucleotides that have at least 70%, preferably at least 75%, at least 80%, at least 85%, at least 90% or at least 95% sequence identity to a CYP3A5 gene, wherein said polynucleotide is having a nucleotide exchange, a nucleotide deletion of at least one nucleotide, or at least one additional nucleotide at a position corresponding to position -20643, -20555, -20359, -20367, -20329, -20323, -20310, -6200, -6177, -4336, -3990, -3868, -3844, -3557, -1617, -795, -86, -74, 136, 174 to 176, 230, 3705, 3709/3710, 5215, 5235, 5516, 7182, 7207, 7303, 7424/7427, 12907, 13028, 13077, 13173, 13226, 13376, 14720, 14836, 14903, 15788, 16079, 16931/16932, 16993, 17163, 19069, 19165, 19208, 27050, 27131/27132, 27526, 31499, 31551 or 31611

of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614).

The term "hybridizing" as used herein refers to polynucleotides which are capable of hybridizing to the polynucleotides of the invention or parts thereof which are associated with a CYP3A5 dysfunction or dysregulation. Thus, said hybridizing polynucleotides are also associated with said dysfunctions and dysregulations. Therefore, said polynucleotides may be useful as probes in Northern or Southern Blot analysis of RNA or DNA preparations, respectively, or can be used as oligonucleotide primers in PCR analysis dependent on their respective size. Also comprised by the invention are hybridizing polynucleotides which are useful for analysing DNA-Protein interactions via, e.g., electrophoretic mobility shift analysis (EMSA). Preferably, said hybridizing polynucleotides comprise at least 10, more preferably at least 15 nucleotides in length while a hybridizing polynucleotide of the present invention to be used as a probe preferably comprises at least 100, more preferably at least 200, or most preferably at least 500 nucleotides in length. It is well known in the art how to perform hybridization experiments with nucleic acid molecules, i.e. the person skilled in the art knows what hybridization conditions s/he has to use in accordance with the present invention. Such hybridization conditions are referred to in standard text books such as Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory (1989) N.Y. Preferred in accordance with the present inventions are polynucleotides which are capable of hybridizing to the polynucleotides of the invention or parts thereof which are associated with a CYP3A5 dysfunction or dysregulation under stringent hybridization conditions, i.e. which do not cross hybridize to unrelated polynucleotides such as polynucleotides encoding a polypeptide different from the CYP3A5 polypeptides of the invention.

Nucleic acid hybridization will be affected by such conditions as salt concentration, temperature, or organic solvents, in addition to the base composition, length of the complementary strands and the number of nucleotide base mismatches between the hybridizing nucleic acids, as will be readily appreciated by those skilled in the art. Stringent temperature conditions will generally include temeratures in excess of 30°C, typically 37°C, and preferably in excess of 45°C. Stringent salt conditions will ordinarily be less than 1000 mM, typically less than 500 mM and preferably less than 200 mM. However, the combination of parameters is much more important than the measure of any single parameter. See, e.g., Wetmur and Davidson, 1968. Probe sequences may also hybridize

specifically to duplex DNA under certain conditions to form triplex or higher order DNA complexes. The preparation of such probes and suitable hybridization conditions are well known in the art.

The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over a strech of at least nine nucleotides, usually at least 20 nucleotides, more usually at least 24 nucleotides, typically at least 28 nucleotides, more typically at least 32 nucleotides, and preferably at least 36 nucleotides or more nucleotides. There are a number of different algorithms known in the art which can be used to measure nucleotide sequence identity. For instance, polynucleotide sequences can be compared using Fasta, a program in GCG Version 6.6, Fasta provides alignments and percent sequence identity of the regions of the best overlap between the query and the search sequence (Pearson, 1980, herein incorporated by reference). For instance, percent sequence identity between nucleic acid sequences can be determined using Fasta with its default parameters (a word size of 6 and the NOPAMfactor for the scoring matrix) as provided in GCG Version 6.1, herein incorporated by reference.

The term "corresponding" as used herein means that a position is not only determined by the number of the preceding nucleotides and amino acids, respectively. The position of a given nucleotide or amino acid in accordance with the present invention which may be deleted, substituted or comprise one or more additional nucleotide(s) may vary due to deletions or additional nucleotides or amino acids elsewhere in the gene or the polypeptide. Thus, under a "corresponding position" in accordance with the present invention it is to be understood that nucleotides or amino acids may differ in the indicated number but may still have similar neighboring nucleotides or amino acids. Said nucleotides or amino acids which may be exchanged, deleted or comprise additional nucleotides or amino acids may for instance together with their neighbors form sequences which may be involved in the regulation of gene expression, stability of the corresponding RNA or RNA editing, as well as encode functional domains or motifs of the protein of the invention.

By, e.g., "position 3709/3710" it is meant that said polynucleotide comprises one or more additional nucleotide(s) which are inserted between positions 3709 and position 3710 of

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the corresponding wild type version of said polynucleotide. The same applies mutatis mutandis to all other position numbers referred to in the above embodiment which are drafted in the same format, i.e. two consecutive position numbers separated by a slash (/). By, e.g., "position 7424 to 7427" is meant that said polynucleotide comprises one or more deleted nucleotides which are deleted between positions 7424 and position 7427 of the corresponding wild type version of said polynucleotide and/or one or more additional nucleotide(s) which are inserted between positions 7424 and position 7427 of the corresponding wild type version of said polynucleotide. The same applies mutatis mutandis to all other position numbers referred to in the above embodiment which are drafted in the same format.

The numbering of the polymorphisms refers to the aligned and joined genomic sequences AF280107.1 and AC005020.2, wherein the T at position 174832 (which has been numbered +8613) of the sequence AF280107.1 refers to position 27340 of the sequence AC005020.2. The nucleotide A at position 27341 of the sequence AC005020.2 has been numbered +8614. Numbering of polymorphisms to a position corresponding to a position up to +8613 refers to the genomic sequence AF280107.1, numbering of polymorphisms to a position corresponding to position +8614 and greater refer to the genomic sequence AC005020.2.

In accordance with the present invention, the mode and population distribution of genetic variations in the CYP3A5 gene has been analyzed by sequence analysis of relevant regions of the human said gene from many different individuals. It is a well known fact that genomic DNA of individuals, which harbor the individual genetic makeup of all genes, including the CYP3A5 gene, can easily be purified from individual blood samples. These individual DNA samples are then used for the analysis of the sequence composition of the alleles of the CYP3A5 gene that are present in the individual which provided the blood sample. The sequence analysis was carried out by PCR amplification of relevant regions of said genes, subsequent purification of the PCR products, followed by automated DNA sequencing with established methods (e.g. ABI dyeterminator cycle sequencing).

One important parameter that had to be considered in the attempt to determine the individual genotypes and identify novel variants of the CYP3A5 gene by direct DNA-sequencing of PCR-products from human blood genomic DNA is the fact that each human narbors (usually, with very few abnormal exceptions) two gene copies of each autosomal gene (diploidy). Because of that, great care had to be taken in the evaluation of the sequences to be able to identify unambiguously not only homozygous sequence variations

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but also heterozygous variations. The details of the different steps in the identification and characterization of novel polymorphisms in the CYP3A5 gene (homozygous and heterozygous) are described in the Examples below.

Over the past 20 years, genetic heterogeneity has been increasingly recognized as a significant source of variation in drug response. Many scientific communications (Meyer, Ann. Rev. Pharmacol. Toxicol. 37 (1997), 269-296 and West, J. Clin. Pharmacol. 37 (1997), 635-648) have clearly shown that some drugs work better or may even be highly toxic in some patients than in others and that these variations in patient's responses to drugs can be related to molecular basis. This "pharmacogenomic" concept spots correlations between responses to drugs and genetic profiles of patient's (Marshall, Nature Biotechnology, 15 (1997), 954-957; Marshall, Nature Biotechnology, 15 (1997), 1249-1252). In this context of population variability with regard to drug therapy, pharmacogenomics has been proposed as a tool useful in the identification and selection of patients which can respond to a particular drug without side effects. This identification/selection can be based upon molecular diagnosis of genetic polymorphisms by genetyping DNA from leukocytes in the blood of patient, for example, and characterization of disease (Bertz, Clin. Pharmacokinet. 32 (1997), 210-256; Engel, J. Chromatogra. B. Biomed. Appl. 678 (1996), 93-103). For the founders of health care, such as health maintenance organizations in the US and government public health services in many European countries, this pharmacogenomics approach can represent a way of both improving health care and reducing overheads because there is a large cost to unnecessary drugs, ineffective drugs and drugs with side effects.

The mutations in the variant genes of the invention sometime result in amino acid deletion(s), insertion(s) and in particular in substitution(s) either alone or in combination. It is of course also possible to genetically engineer such mutations in wild type genes or other mutant forms. Methods for introducing such modifications in the DNA sequence of said genes are well known to the person skilled in the art; see, e.g., Sambrook, Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory (1989) N.Y.

For the investigation of the nature of the alterations in the amino acid sequence of the polypeptides of the invention software programs may be used such as RASMOL that are obtainable from the Internet. Furthermore, folding simulations and computer redesign of structural motifs can be performed using other appropriate computer programs (Olszewski,

Proteins 25 (1996), 286-299; Hoffman, Comput. Appl. Biosci. 11 (1995), 675-679). Computers can be used for the conformational and energetic analysis of detailed protein models (Monge, J. Mol. Biol. 247 (1995), 995-1012; Renouf, Adv. Exp. Med. Biol. 376 (1995), 37-45). These analysis can be used for the identification of the influence of a particular mutation on binding and/or processing of drugs.

Usually, said amino acid deletion, addition or substitution in the amino acid sequence of the protein encoded by the polynucleotide of the invention is due to one or more nucleotide substitution, insertion or deletion, or any combinations thereof. Preferably said nucleotide substitution, insertion or deletion may result in amino acid substitutions of HGLFK to YGTF. (with the period meaning termination) at a position corresponding to position 30 to 34 of the CYP3A5 polypeptide (Accession No: NP_000768.1), in an amino acid substitution of S to Y at a position corresponding to position 100 of the CYP3A5 polypeptide (Accession No: NP_000768.1), in amino acid substitutions of TYD to YL. (with the period meaning termination) at a position corresponding to position 346 to 348 of the CYP3A5 polypeptide (Accession No: NP_000768.1), or in an amino acid substitution of T to N at a position corresponding to position 398 of the CYP3A5 polypeptide (Accession No: NP_000768.1).

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The mutations in the CYP3A5 gene detected in accordance with the present invention are listed in Table 2A-E. The methods of the mutation analysis followed standard protocols and are described in detail in the Examples. In general such methods are to be used in accordance with the present invention for evaluating the phenotypic spectrum as well as the overlapping clinical characteristics of diseases or conditions related to dysfunctions and diseases related to the drug metabolism. Advantageously, the characterization of said mutants may form the basis of the development of improved drugs, such as drugs which are used e.g. in cancer therapy and diseases including cardiovascular diseases, diabetes and AIDS. Said methods encompass for example haplotype analysis, single-strand conformation polymorphism analysis (SSCA), PCR and direct sequencing, or TaqMan® analysis. On the basis of thorough clinical characterization of many patients the phenotypes can then be correlated to these mutations as well as to mutations that had been described earlier, for example in Jounaidi, Biochem Biophys Res Commun, 221, pp. 466-470, 1996.

Also comprised by the polynucleotides referred to in the present invention are polynucleotides which comprise at least two, preferably at least three, of the polynucleotides specified hereinabove, i.e. polynucleotides having a nucleotide sequence which contains at least two, preferably three of the mutations comprised by the above polynucleotides or listed in the tables below. Thus, the haplotype determined in accordance with the present invention can be characterized by at least two, preferably three of said mutations in the CYP3A5 locus. Further, the polynucleotide of the invention may further comprise at least one nucleotide deletion, addition and/or substitution other than those specified hereinabove, for example those described in the prior art; e.g., in Jounaidi, Biochem Biophys Res Commun, 221, pp. 466-470, 1996, in Paulussen, Pharmacogenetics 10, pp. 415-424, 2000, in Kuehl, 2001, Nature Genetics 27: 383-391, or in Chou, 2001, Drug Metab Dispos 29: 1205-1209.

This allows the study of synergistic effects of said mutations in the CYP3A5 gene and/or a polypeptide encoded by said polynucleotide on the pharmacological profile of drugs in patients who bear such mutant forms of the gene or similar mutant forms that can be mimicked by the above described proteins. It is expected that the analysis of said synergistic effects provides deeper insights into the onset of dysfunctions or diseases related to drug metabolism as described supra. From said deeper insight the development of diagnostic and pharmaceutical compositions related to dysfunctions or diseases related to drug metabolism will greatly benefit.

Moreover, it has been surprisingly found that the so called positive predictive power for CYP3A5 dysfunctions or dysregulations can be significantly increased based on the polynucleotides of the present invention and thus allows a reliable prediction in contrast to positive predictive power based on the prior art. The increased CYP3A5 protein expression in all except one liver samples (17/18) identified in accordance with the present invention and described in detail in the examples below co-segregates with a haplotype which consists of at least three variants (ch-v-021, ch-v-026, ch-v-015) with distinct locations within or upstream of the gene locus. Genotyping these three variants has in no case led to the generation of false-positive predictions resulting in an estimated positive predictive power for the 3-variant genotype of about 99.95 %. This is in striking contrast to the positive predictive power determined for the haplotype described by Paulussen, Pharmacogenetics 10, pp. 415-424, 2000 which is about 65 %. Moreover, based on the polynucleotides of the invention and as described in the examples below, it has been found that the SNPs described by Paulussen, Pharmacogenetics 10, pp. 415-424, 2000

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are located in contrary to what is reported in said document approximately 20 kb upstream of the transcriptional start site of the CYP3A5 gene in a sequence 5' to a CYP3A5 pseudogene locus.

Therefore, the haplotypes characterized on the basis of the polynucleotides of the present invention fulfil the criteria expected from a reliable marker of CYP3A5 expression. As is evident to the person skilled in the art, the genetic knowledge deduced from the present invention can now be used to exactly and reliably characterize the genotype of a patient. Advantageously, diseases or a prevalence for a disease which are associated with CYP3A5 dysfunction or dysregulation, such as cancer, diseases including cardiovascular diseases, diabetes and AIDS, can be predicted and preventive or therapeutical measures can be applied accordingly. Moreover in accordance with the foregoing, in cases where a given drug takes an unusual effect, a suitable individual therapy can be designed based on the knowledge of the individual genetic makeup of a subject with respect to the polynucleotides of the invention and improved therapeutics can be developed as will be further discussed below.

Finally, the polynucleotides and polypeptides referred to in accordance with the present invention are also useful as forensic markers, which improve the identification of subjects which have been murdered or killed by, for example, a crime of violence or any other violence and can not be identified by the well known conventional forensic methods. The application of forensic methods based on the detection of the polymorphisms comprised by the polynucleotides of this invention in the genome of a subject are particularly well suited in cases where a (dead) body is disfigured in a severe manner such that identification by other body characteristics such as the features of the face is not possible. This is the case, for example, for corpse found in water which are usually entirely disfigured. Advantageously methods which are based on the provision of the polynucleotides of the invention merely require a minimal amount of tissues or cells in order to be carried out. Said tissues or cells may be blood droplets, hair roots, epidermal scales, saliva droplets, sperms etc. Since only such a minimal amount of tissues or cells are required for the identification of a subject, the polymorphisms comprised by the polynucleotides of this invention can be also used as forensic markers in order to prove someone guilt of a crime, such as a violation or a ravishment. Moreover, the polymorphisms comprised by the polynucleotides of this invention can be used to proof paternity. In accordance with the forensic methods referred to herein the presence or absence of the polynucleotides of the invention is determined and compared with a reference sample which is unambiguously derived from the subject to be identified. The

forensic methods which require detection of the presence or absence of the polynucleotides of the invention in a sample of a subject the polymorphisms comprised by the polynucleotides of this invention can be for example PCR-based techniques which are particularly well suited in cases where only a minimal amount of tissues or cells are available as forensic samples. On the other hand, where enough tissue or cells are available, hybridization based techniques may be performed in order to detect the presence or absence of a polynucleotide of this invention. These techniques are well known by the person skilled in the art and can be adopted to the individual purposes referred to herein without further ado. In conclusion, thanks to the present invention forensic means which allow improved and reliable predictions as regards the aforementioned aspects are now available.

In line with the foregoing, preferably, the polynucleotide of the present invention is associated with cancer or diseases including cardiovascular diseases, diabetes and AIDS.

The term "cancer" used herein is very well known and characterized in the art. Several variants of cancer exist and are comprised by said term as meant in accordance with the invention. For a detailed list of symptoms which are indicative for cancer it is referred to text book knowledge, e.g. Pschyrembel.

In a further embodiment the present invention relates to a polynucleotide which is DNA or RNA.

The polynucleotide of the invention may be, e.g., DNA, cDNA, genomic DNA, RNA or synthetically produced DNA or RNA or a recombinantly produced chimeric nucleic acid molecule comprising any of those polynucleotides either alone or in combination. Preferably said polynucleotide is part of a vector, particularly plasmids, cosmids, viruses and bacteriophages used conventionally in genetic engineering that comprise a polynucleotide of the invention. Such vectors may comprise further genes such as marker genes which allow for the selection of said vector in a suitable host cell and under suitable conditions.

The invention furthermore relates to a gene comprising the polynucleotide of the invention.

It is well known in the art that genes comprise structural elements which encode an amino acid sequence as well as regulatory elements which are involved in the regulation of the expression of said genes. Structural elements are represented by exons which may either encode an amino acid sequence or which may encode for RNA which is not encoding an amino acid sequence but is nevertheless involved in RNA function, e.g. by regulating the stability of the RNA or the nuclear export of the RNA.

Regulatory elements of a gene may comprise promoter elements or enhancer elements both of which could be involved in transcriptional control of gene expression. It is very well known in the art that a promoter is to be found upstream of the structural elements of a gene. Regulatory elements such as enhancer elements, however, may be found distributed over the entire locus of the gene. Said elements could be reside, e.g., in introns, regions of genomic DNA which separate the exons of a gene. Said introns may comprise further regulatory elements which are required for proper gene expression. Introns are usually transcribed together with the exons of a gene resulting in a nascent RNA transcript which contains both, exon and intron sequences. The intron encoded RNA sequences are usually removed by a process known as RNA splicing. However, said process also requires regulatory sequences present on a RNA transcript, said regulatory sequences may be encoded by the introns.

In addition, besides their function in transcriptional control and control of proper RNA processing and/or stability, regulatory elements of a gene could be also involved in the control of genetic stability of a gene locus. Said elements control, e.g., recombination events or serve to maintain a certain structure of the DNA or the arrangement of DNA in a chromosome.

Therefore, polymorphisms can occur in exons of a gene which encode an amino acid sequence as discussed supra as well as in regulatory regions which are involved in the above discussed process. The analysis of the nucleotide sequence of a gene locus in its entirety including, e.g., introns is in light of the above desirable. It has been found based on the polymorphisms comprised by the polynucleotides of the present invention that the mechanism of the increased expression of CYP3A5 protein in most Caucasians livers described in the examples below may involve enhanced transcription and stabilisation of the gene's transcripts.

Therefore, in a furthermore preferred embodiment of the gene of the invention a nucleotide deletion, addition and/or substitution results in altered expression of the variant gene compared to the corresponding wild type gene.

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In another embodiment the present invention relates to a vector comprising the polynucleotide of the invention or the gene of the invention.

Said vector may be, for example, a phage, plasmid, viral or retroviral vector. Retroviral vectors may be replication competent or replication defective. In the latter case, viral propagation generally will occur only in complementing host/cells.

The polynucleotides or genes of the invention may be joined to a vector containing selectable markers for propagation in a host. Generally, a plasmid vector is introduced in a precipitate such as a calcium phosphate precipitate, or in a complex with a charged lipid or in carbon-based clusters. Should the vector be a virus, it may be packaged in vitro using an appropriate packaging cell line prior to application to host cells.

In a more preferred embodiment of the vector of the invention the polynucleotide is operatively linked to expression control sequences allowing expression in prokaryotic or aukaryotic cells or isolated fractions thereof.

Expression of said polynucleotide comprises transcription of the polynucleotide, preferably into a translatable mRNA. Regulatory elements ensuring expression in eukaryotic cells, preferably marnmalian cells, are well known to those skilled in the art. They usually comprise regulatory sequences ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript. Additional regulatory elements may include transcriptional as well as translational enhancers. Possible regulatory elements permitting expression in prokaryotic host cells comprise, e.g., the lac, trp or tac promoter in E. coli, and examples for regulatory elements permitting expression in eukaryotic host cells are the AOX1 or GAL1 promoter in yeast or the CMV-, SV40-, RSV-promoter (Rous sarcoma virus), CMV-enhancer, SV40-enhancer or a globin intron in mammalian and other animal cells. Beside elements which are responsible for the initiation of transcription such regulatory elements may also comprise transcription termination signals, such as the SV40-poly-A site or the tk-poly-A site, downstream of the polynucleotide. In this context, suitable expression vectors are known in the art such as Okayama-Berg cDNA expression vector pcDV1 (Pharmacia), pCDM8, pRc/CMV, pcDNA1, pcDNA3 (In-vitrogene), pSPORT1 (GIBCO BRL). Preferably, said vector is an expression vector and/or a gene transfer or targeting vector. Expression vectors derived from viruses such as retroviruses, vaccinia virus, adeno-associated virus, herpes viruses, or bovine

papilloma virus, may be used for delivery of the polynucleotides or vector of the invention into targeted cell population. Methods which are well known to those skilled in the art can be used to construct recombinant viral vectors; see, for example, the techniques described in Sambrook, Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory (1989) N.Y. and Ausubel, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y. (1994). Alternatively, the polynucleotides and vectors of the invention can be reconstituted into liposomes for delivery to target cells.

The term "isolated fractions thereof" refers to fractions of eukaryotic or prokaryotic cells or tissues comprising said cells which are capable of transcribing or transcribing and translating RNA from the vector of the invention. Said fractions comprise proteins which are required for transcrition of RNA or transcription of RNA and translation of said RNA into a polypeptide. Said isolated fractions may be, e.g., nuclear and cytoplasmic fractions of eukaryotic cells such as of reticulocytes.

The present invention furthermore relates to a host cell genetically engineered with the polynucleotide of the invention, the gene of the invention or the vector of the invention.

Said host cell may be a prokaryotic or eukaryotic cell; see supra. The polynucleotide or vector of the invention which is present in the host cell may either be integrated into the genome of the host cell or it may be maintained extrachromosomally. In this respect, it is also to be understood that the recombinant DNA molecule of the invention can be used for "gene targeting" and/or "gene replacement", for restoring a mutant gene or for creating a mutant gene via homologous recombination; see for example Mouellic, Proc. Natl. Acad. Sci. USA, 87 (1990), 4712-4716; Joyner, Gene Targeting, A Practical Approach, Oxford University Press.

The host cell can be any prokaryotic or eukaryotic cell, such as a bacterial, insect, fungal, plant, animal or human cell. Preferred fungal cells are, for example, those of the genus Saccharomyces, in particular those of the species S. cerevisiae. The term "prokaryotic" is meant to include all bacteria which can be transformed or transfected with a polynucleotide for the expression of a variant polypeptide of the invention. Prokaryotic hosts may include gram negative as well as gram positive bacteria such as, for example, E. coli, S. typhimurium, Serratia marcescens and Bacillus subtilis. A polynucleotide coding for a mutant form of variant polypeptides of the invention can be used to transform or transfect the host using any of the techniques commonly known to those of ordinary skill in the art.

Methods for preparing fused, operably linked genes and expressing them in bacteria or animal cells are well-known in the art (Sambrook, supra). The genetic constructs and methods described therein can be utilized for expression of variant polypeptides of the invention in, e.g., prokaryotic hosts. In general, expression vectors containing promoter sequences which facilitate the efficient transcription of the inserted polynucleotide are used in connection with the host. The expression vector typically contains an origin of replication, a promoter, and a terminator, as well as specific genes which are capable of providing phenotypic selection of the transformed cells. The transformed prokaryotic hosts can be grown in fermentors and cultured according to techniques known in the art to achieve optimal cell growth. The proteins of the invention can then be isolated from the grown medium, cellular lysates, or cellular membrane fractions. The isolation and purification of the microbially or otherwise expressed polypeptides of the invention may be by any conventional means such as, for example, preparative chromatographic separations and immunological separations such as those involving the use of monoclonal or polyclonal antibodies.

Thus, in a further embodiment the invention relates to a method for producing a molecular variant polypeptide or fragment thereof comprising culturing the above described host cell; and recovering said protein or fragment from the culture.

in another embodiment the present invention relates to a method for producing cells capable of expressing a molecular variant polypeptide comprising genetically engineering cells with the polynucleotide of the invention, the gene of the invention or the vector of the invention.

The cells obtainable by the method of the invention can be used, for example, to test drugs according to the methods described in D. L. Spector, R. D. Goldman, L. A. Leinwand, Cells, a Lab manual, CSH Press 1998. Furthermore, the cells can be used to study known drugs and unknown derivatives thereof for their ability to complement the deficiency caused by mutations in the CYP3A5 gene. For these embodiments the host cells preferably lack a wild type allele, preferably both alleles of the CYP3A5 gene and/or have at least one mutated from thereof. Ideally, the gene comprising an allele as comprised by the polynucleotides of the invention could be introduced into the wild type locus by homologous replacement. Alternatively, strong overexpression of a mutated allele over the normal allele and comparison with a recombinant cell line overexpressing the normal

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allele at a similar level may be used as a screening and analysis system. The cells obtainable by the above-described method may also be used for the screening methods referred to herein below.

Furthermore, the invention relates to a polypeptide or fragment thereof encoded by the polynucleotide of the invention, the gene of the invention or obtainable by the method described above or from cells produced by the method described above.

In this context it is also understood that the variant polypeptide of the invention can be further modified by conventional methods known in the art. By providing said variant proteins according to the present invention it is also possible to determine the portions relevant for their biological activity or inhibition of the same. The terms "polypeptide" and "protein" as used herein are exchangeable. Moreover, what is comprised by said terms is standard textbook knowledge.

The present invention furthermore relates to an antibody which binds specifically to the polypeptide of the invention.

Advantageously, the antibody specifically recognizes or binds an epitope containing one or more amino acid substitution(s) as defined above. Antibodies against the variant polypeptides of the invention can be prepared by well known methods using a purified protein according to the invention or a (synthetic) fragment derived therefrom as an antigen. Monoclonal antibodies can be prepared, for example, by the techniques as originally described in Köhler and Milstein, Nature 256 (1975), 495, and Galfré, Meth. Enzymol. 73 (1981), 3, which comprise the fusion of mouse myeloma cells to spleen cells derived from immunized mammals. In a preferred embodiment of the invention, said antibody is a monoclonal antibody, a polyclonal antibody, a single chain antibody, human or humanized antibody, primatized, chimerized or fragment thereof that specifically binds said peptide or polypeptide also including bispecific antibody, synthetic antibody, antibody fragment, such as Fab, Fv or scFv fragments etc., or a chemically modified derivative of any of these. Furthermore, antibodies or fragments thereof to the aforementioned polypeptides can be obtained by using methods which are described, e.g., in Harlow and Lane "Antibodies, A Laboratory Manual", CSH Press, Cold Spring Harbor, 1988. These antibodies can be used, for example, for the immunoprecipitation and immunolocalization of the variant polypeptides of the invention as well as for the monitoring of the presence of

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said variant polypeptides, for example, in recombinant organisms, and for the identification of compounds interacting with the proteins according to the invention. For example, surface plasmon resonance as employed in the BIAcore system can be used to increase the efficiency of phage antibodies which bind to an epitope of the protein of the invention (Schier, Human Antibodies Hybridomas 7 (1996), 97-105; Malmborg, J. Immunol. Methods 183 (1995), 7-13).

In a preferred embodiment the antibody of the present invention specifically recognizes an epitope containing one or more amino acid substitution(s) resulting from a nucleotide exchange as defined supra.

Antibodies which specifically recognize modified amino acids such as phospho-Tyrosine residues are well known in the art. Similarly, in accordance with the present invention antibodies which specifically recognize even a single amino acid exchange in an epitope may be generated by the well known methods described supra.

In light of the foregoing, in a more preferred embodiment the antibody of the present invention is monoclonal or polyclonal.

The invention also relates to a transgenic non-human animal comprising at least one polynucleotide of the invention, the gene of the invention or the vector of the invention as described supra.

The present invention also encompasses a method for the production of a transgenic nonhuman animal comprising introduction of a polynucleotide or vector of the invention into a
germ cell, an embryonic cell, stem cell or an egg or a cell derived therefrom. The nonhuman animal can be used in accordance with the method of the invention described
below and may be a non-transgenic healthy animal, or may have a disease or disorder,
preferably a disease caused by at least one mutation in the gene of the invention. Such
transgenic animals are well suited for, e.g., pharmacological studies of drugs in connection
with variant forms of the above described variant polypeptides since these polypeptides or
at least their functional domains are conserved between species in higher eukaryotes,
particularly in mammals. Production of transgenic embryos and screening of those can be
performed, e.g., as described by A. L. Joyner Ed., Gene Targeting, A Practical Approach

(1993), Oxford University Press. The DNA of the embryos can be analyzed using, e.g., Southern blots with an appropriate probe or based on PCR techniques.

A transgenic non-human animal in accordance with the invention may be a transgenic mouse, rat, hamster, dog, monkey, rabbit, pig, frog, nematode such as Caenorhabditis elegans, fruitfly such as Drosophila melanogaster or fish such as torpedo fish or zebrafish comprising a polynucleotide or vector of the invention or obtained by the method described above, preferably wherein said polynucleotide or vector is stably integrated into the genome of said non-human animal, preferably such that the presence of said polynucleotide or vector leads to the expression of the variant polypeptide of the invention. It may comprise one or several copies of the same or different polynucleotides or genes of the invention. This animal has numerous utilities, including as a research model for cancer or diseases including cardiovascular diseases, diabetes and AIDS or any other disease caused by as dysfunction or dysregulation of the polynucleotides or polypeptides of the invention research and therefore, presents a novel and valuable animal in the development of therapies, treatment, etc. for cancer diseases or diseases including cardiovascular diseases, diabetes and AIDS or any other disease caused by as dysfunction or dysregulation of the polynucleotides or polypeptides of the invention. Accordingly, in this instance, the mammal is preferably a laboratory animal such as a mouse or rat.

Thus, in a preferred embodiment the transgenic non-human animal of the invention is a mouse, a rat or a zebrafish.

Numerous reports revealed that said animals are particularly well suited as model organisms for the investigation of the drug metabolism and its deficiencies or cancer. Advantageously, transgenic animals can be easily created using said model organisms, due to the availability of various suitable techniques well known in the art.

The invention also relates to a solid support comprising one or a plurality of the polynucleotide, the gene, the vector, the polypeptide, the antibody or the host cell of the invention in immobilized form.

The term "solid support" as used herein refers to a flexible or non-flexible support that is suitable for carrying said immobilized targets. Said solid support may be homogenous or

inhomogeneous. For example, said solid support may consist of different materials having the same or different properties with respect to flexibility and immobilization, for instance, or said solid support may consist of one material exhibiting a plurality of properties also comprising flexibility and immobilization properties. Said solid support may comprise glass-, polypropylene- or silicon-chips, membranes, oligonucleotide-conjugated beads or bead arrays.

The term "immobilized" means that the molecular species of interest is fixed to a solid support, preferably covalently linked thereto. This covalent linkage can be achieved by different means depending on the molecular nature of the molecular species. Moreover, the molecular species may be also fixed on the solid support by electrostatic forces, hydrophobic or hydrophilic interactions or Van-der-Waals forces. The above described physico-chemical interactions typically occur in interactions between molecules. For example, biotinylated polypeptides may be fixed on a avidin-coated solid support due to interactions of the above described types. Further, polypeptides such as antibodies, may be fixed on an antibody coated solid support. Moreover, the immobilization is dependent on the chemical properties of the solid support. For example, the nucleic acid molecules can be immobilized on a membrane by standard techniques such as UV-crosslinking or heat.

in a preferred embodiment of the invention said solid support is a membrane, a glass- or poylpropylene- or silicon-chip, are oligonucleotide-conjugated beads or a bead array, which is assembled on an optical filter substrate.

Moreover, the present invention relates to an in vitro method for identifying a polymorphism said method comprising the steps of:

- (a) isolating a polynucleotide or the gene of the invention from a plurality of subgroups of individuals, wherein one subgroup has no prevalence for a CYP3A5 associated disease and at least one or more further subgroup(s) do have prevalence for a CYP3A5 associated disease; and
- (b) identifying a polymorphism by comparing the nucleic acid sequence of said polynucleotide or said gene of said one subgroup having no prevalence for a CYP3A5 associated disease with said at least one or more further subgroup(s) having a prevalence for a CYP3A5 associated disease.

The term "prevalence" as used herein means that individuals are susceptible for one or more disease(s) which are associated with CYP3A5 dysfuntion or dysregulation or could already have one or more of said disease(s). Thereby, one CYP3A5 associated disease can be used to determine the susceptibility for another CYP3A5 associated disease, e.g. impaired drug metabolism may be indicative for a prevalence for, e.g. cancer. Moreover, symptoms which are indicative for a prevalence for developing said diseases are very well known in the art and have been sufficiently described in standard textbooks such as Pschyrembel.

Advantageously, polymorphisms according to the present invention which are associated with CYP3A5 dysfunction or dysregulation or one or more disease(s) based thereon should be enriched in subgroups of individuals which have a prevalence for said diseases versus subgroups which have no prevalence for said diseases. Thus, the above described method allows the rapid and reliable detection of polymorphisms which are indicative for one or more CYP3A5 associated disease(s) or a susceptibility therefor. Advantageously, due to the phenotypic preselection a large number of individuals having no prevalence might be screened for polymorphisms in general. Thereby, a reference sequences comprising polymorphisms which do not correlate to one or more CYP3A5 associated disease(s) can be obtained. Based on said reference sequences it is possible to efficiently and reliably determine the relevant polymorphisms.

In a further embodiment the present invention relates to a method for identifying and obtaining a pro-drug or a drug capable of modulating the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:

- (a) contacting the polypeptide, the solid support of the invention, a cell expressing a molecular variant gene comprising a polynucleotide of the invention, the gene or the vector of the invention in the presence of components capable of providing a detectable signal in response to drug activity with a compound to be screened for pro-drug or drug activity; and
- (b) detecting the presence or absence of a signal or increase or decrease of a signal generated from the pro-drug or the drug activity, wherein the absence, presence, increase or decrease of the signal is indicative for a putative pro-drug or drug.

The term "compound" in a method of the invention includes a single substance or a plurality of substances which may or may not be identical.

Said compound(s) may be chemically synthesized or produced via microbial fermentation but can also be comprised in, for example, samples, e.g., cell extracts from, e.g., plants, animals or microorganisms. Furthermore, said compounds may be known in the art but hitherto not known to be useful as an inhibitor, respectively. The plurality of compounds may be, e.g., added to the culture medium or injected into a cell or non-human animal of the invention.

If a sample containing (a) compound(s) is identified in the method of the invention, then it is either possible to isolate the compound from the original sample identified as containing the compound in question or one can further subdivide the original sample, for example, if it consists of a plurality of different compounds, so as to reduce the number of different substances per sample and repeat the method with the subdivisions of the original sample. It can then be determined whether said sample or compound displays the desired properties, for example, by the methods described herein or in the literature (Spector et al., Cells manual; see supra). Depending on the complexity of the samples, the steps described above can be performed several times, preferably until the sample identified according to the method of the invention only comprises a limited number of or only one substance(s). Preferably said sample comprises substances of similar chemical and/or physical properties, and most preferably said substances are identical. The methods of the present invention can be easily performed and designed by the person skilled in the art, for example in accordance with other cell based assays described in the prior art or by using and modifying the methods as described herein. Furthermore, the person skilled in the art will readily recognize which further compounds may be used in order to perform the methods of the invention, for example, enzymes, if necessary, that convert a certain compound into a precursor. Such adaptation of the method of the invention is well within the skill of the person skilled in the art and can be performed without undue experimentation.

Compounds which can be used in accordance with the present invention include peptides, proteins, nucleic acids, antibodies, small organic compounds, ligands, peptidomimetics, PNAs and the like. Said compounds may act as agonists or antagonists of the invention. Said compounds can also be functional derivatives or analogues of known drugs. Methods for the preparation of chemical derivatives and analogues are well known to those skilled in the art and are described in, for example, Beilstein, Handbook of Organic Chemistry, Springer edition New York Inc., 175 Fifth Avenue, New York, N.Y. 10010 U.S.A. and Organic Synthesis, Wiley, New York, USA. Furthermore, said derivatives and analogues

can be tested for their effects according to methods known in the art or as described. Furthermore, peptide mimetics and/or computer aided design of appropriate drug derivatives and analogues can be used, for example, according to the methods described below. Such analogs comprise molecules may have as the basis structure of known CYP3A5 substrates and/or inhibitors and/or modulators; see infra.

Appropriate computer programs can be used for the identification of interactive sites of a putative inhibitor and the polypeptides of the invention by computer assistant searches for complementary structural motifs (Fassina, Immunomethods 5 (1994), 114-120). Further appropriate computer systems for the computer aided design of protein and peptides are described in the prior art, for example, in Berry, Biochem. Soc. Trans. 22 (1994), 1033-1036; Wodak, Ann. N. Y. Acad. Sci. 501 (1987), 1-13; Pabo, Biochemistry 25 (1986), 5987-5991. The results obtained from the above-described computer analysis can be used in combination with the method of the invention for, e.g., optimizing known inhibitors, analogs, antagonists or agonists. Appropriate peptidomimetics and other inhibitors can also be identified by the synthesis of peptidomimetic combinatorial libraries through successive chemical modification and testing the resulting compounds, e.g., according to the methods described herein. Methods for the generation and use of peptidomimetic combinatorial libraries are described in the prior art, for example in Ostresh, Methods in Enzymology 267 (1996), 220-234 and Dorner, Bioorg. Med. Chem. 4-(1996), 709-715. Furthermore, the three-dimensional and/or crystallographic structure of said compounds and the polypeptides of the invention can be used for the design of peptidomimetic drugs (Rose, Biochemistry 35 (1996), 12933-12944; Rutenber, Bioorg. Med. Chem. 4 (1996), 1545-1558). It is very well known how to obtain said compounds, e.g. by chemical or biochemical standard techniques. Thus, also comprised by the method of the invention are means of making or producing said compounds. In summary, the present invention provides methods for identifying and obtaining compounds which can be used in specific doses for the treatment of specific forms of CYP3A5 associated diseases, e.g. dysfunctions of the drug metabolism or cancer.

The above definitions apply mutatis mutandis to all of the methods described in the following.

In a further embodiment the present invention relates to a method for identifying and obtaining an inhibitor of the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:

(a) contacting the protein, the solid support of the invention or a cell expressing a molecular variant gene comprising a polynucleotide or the gene or the vector of the invention in the presence of components capable of providing a detectable signal in response to drug activity with a compound to be screened for inhibiting activity; and

(b) detecting the presence or absence of a signal or increase or decrease of a signal generated from the inhibiting activity, wherein the absence or decrease of the signal is indicative for a putative inhibitor.

In a preferred embodiment of the method of the invention said cell is a cell, obtained by the method of the invention or can be obtained from the transgenic non-human animal as described supra.

In a still further embodiment the present invention relates to a method of identifying and obtaining a pro-drug or drug capable of modulating the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:

- (a) contacting the host cell, the cell obtained by the method of the invention, the polypeptide or the solid support of the invention with the first molecule known to be bound by a CYP3A5 polypeptide to form a first complex of said polypeptide and said first molecule;
- (a) contacting said first complex with a compound to be screened, and
- (c) measuring whether said compound displaces said first molecule from said first complex.

Advantageously, in said method said measuring step comprises measuring the formation of a second complex of said protein and said inhibitor candidate. Preferably, said measuring step comprises measuring the amount of said first molecule that is not bound to said protein.

in a particularly preferred embodiment of the above-described method of said first molecule is a agonist or antagonist or a substrate and/or a inhibitor and/or a modulator of the polypeptide of the invention, e.g., with a radioactive or fluorescent label.

In a still another embodiment the present invention relates to a method of identifying and obtaining an inhibitor capable of modulating the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:

(a) contacting the host cell or the cell obtained by the method of the invention, the protein or the solid support of the invention with the first molecule known to be bound by a CYP3A5 polypeptide to form a first complex of said protein and said first molecule;

- (b) contacting said first complex with a compound to be screened, and
- (c) measuring whether said compound displaces said first molecule from said first complex.

In a preferred embodiment of the method of the invention said measuring step comprises measuring the formation of a second complex of said protein and said compound.

In another preferred embodiment of the method of the invention said measuring step comprises measuring the amount of said first molecule that is not bound to said protein.

In a more preferred embodiment of the method of the invention said first molecule is labeled.

The invention furthermore relates to a method for the production of a pharmaceutical composition comprising the steps of the method as described supra; and the further step of formulating the compound identified and obtained or a derivative thereof in a pharmaceutically acceptable form.

The therapeutically useful compounds identified according to the methods of the invention can be formulated and administered to a patient as discussed above. For uses and therapeutic doses determined to be appropriate by one skilled in the art and for definitions of the term "pharmaceutical composition" see infra.

Furthermore, the present invention encompasses a method for the preparation of a pharmaceutical composition comprising the steps of the above-described methods; and formulating a drug or pro-drug in the form suitable for therapeutic application and preventing or ameliorating the disorder of the subject diagnosed in the method of the invention.

Drugs or pro-drugs after their in vivo administration are metabolized in order to be eliminated either by excretion or by metabolism to one or more active or inactive

metabolites (Meyer, J. Pharmacokinet. Biopharm. 24 (1996), 449-459). Thus, rather than using the actual compound or inhibitor identified and obtained in accordance with the methods of the present invention a corresponding formulation as a pro-drug can be used which is converted into its active in the patient. Precautionary measures that may be taken for the application of pro-drugs and drugs are described in the literature; see, for review, Ozama, J. Toxicol. Sci. 21 (1996), 323-329).

In a preferred embodiment of the method of the present invention said drug or prodrug is a derivative of a medicament as defined hereinafter.

The present invention also relates to a method of diagnosing a disorder related to the presence of a molecular variant of the CYP3A5 gene or susceptibility to such a disorder comprising determining the presence of a polynucleotide or the gene of the invention in a sample from a subject.

In accordance with this embodiment of the present invention, the method of testing the status of a disorder or susceptibility to such a disorder can be effected by using a polynucleotide gene or nucleic acid of the invention, e.g., in the form of a Southern or Northern blot or *in situ* analysis. Said nucleic acid sequence may hybridize to a coding region of either of the genes or to a non-coding region, e.g. intron. In the case that a complementary sequence is employed in the method of the invention, said nucleic acid molecule can again be used in Northern blots. Additionally, said testing can be done in conjunction with an actual blocking, e.g., of the transcription of the gene and thus is expected to have therapeutic relevance. Furthermore, a primer or oligonucleotide can also be used for hybridizing to one of the above mentioned CYP3A5 gene or corresponding mRNAs. The nucleic acids used for hybridization can, of course, be conveniently labeled by incorporating or attaching, e.g., a radioactive or other marker. Such markers are well known in the art. The labeling of said nucleic acid molecules can be effected by conventional methods.

Additionally, the presence or expression of variant CYP3A5 gene can be monitored by using a primer pair that specifically hybridizes to either of the corresponding nucleic acid sequences and by carrying out a PCR reaction according to standard procedures. Specific hybridization of the above mentioned probes or primers preferably occurs at stringent hybridization conditions. The term "stringent hybridization conditions" is well known in the art; see, for example, Sambrook et al., "Molecular Cloning, A Laboratory Manual" second

ed., CSH Press, Cold Spring Harbor, 1989; "Nucleic Acid Hybridisation, A Practical Approach", Hames and Higgins eds., IRL Press, Oxford, 1985. Furthermore, the mRNA, cRNA, cDNA or genomic DNA obtained from the subject may be sequenced to identify mutations which may be characteristic fingerprints of mutations in the polynucleotide or the gene of the invention. The present invention further comprises methods wherein such a fingerprint may be generated by RFLPs of DNA or RNA obtained from the subject, optionally the DNA or RNA may be amplified prior to analysis, the methods of which are well known in the art. RNA fingerprints may be performed by, for example, digesting an RNA sample obtained from the subject with a suitable RNA-Enzyme, for example RNase T₁, RNase T₂ or the like or a ribozyme and, for example, electrophoretically separating and detecting the RNA fragments as described above.

Further modifications of the above-mentioned embodiment of the invention can be easily devised by the person skilled in the art, without any undue experimentation from this disclosure; see, e.g., the examples. An additional embodiment of the present invention relates to a method wherein said determination is effected by employing an antibody of the invention or fragment thereof. The antibody used in the method of the invention may be labeled with detectable tags such as a histidine flags or a biotin molecule.

The invention relates to a method of diagnosing a disorder related to the presence of a molecular variant of a CYP3A5 gene or susceptibility to such a disorder comprising determining the presence of a polypeptide or the antibody of the invention in a sample from a subject.

In a preferred embodiment of the above described method said disorder is cancer or diseases including cardiovascular diseases, diabetes and AIDS.

In a preferred embodiment of the present invention, the above described method is comprising PCR, ligase chain reaction, restriction digestion, direct sequencing, nucleic acid amplification techniques, hybridization techniques or immunoassays.

Said techniques are very well known in the art.

Moreover, the invention relates to a method of detection of the polynucleotide or the gene of the invention in a sample comprising the steps of

 (a) contacting the solid support described supra with the sample under conditions allowing interaction of the polynucleotide or the gene of the invention with the immobilized targets on a solid support and;

(b) determining the binding of said polynucleotide or said gene to said immobilized targets on a solid support.

The invention also relates to an in vitro method for diagnosing a disease comprising the steps of the method described supra, wherein binding of said polynucleotide or gene to said immobilized targets on said solid support is indicative for the presence or the absence of said disease or a prevalence for said disease.

The invention furthermore relates to a diagnostic composition comprising the polynucleotide, the gene, the vector, the polypeptide or the antibody of the invention.

In addition, the invention relates to a pharmaceutical composition comprising the polynucleotide, the gene, the vector, the polypeptide or the antibody of the invention.

These pharmaceutical compositions comprising, e.g., the antibody may conveniently be administered by any of the routes conventionally used for drug administration, for instance, orally, topically, parenterally or by inhalation. Acceptable salts comprise acetate, methylester, HCl, sulfate, chloride and the like. The compounds may be administered in conventional dosage forms prepared by combining the drugs with standard pharmaceutical carriers according to conventional procedures. These procedures may involve mixing, granulating and compressing or dissolving the ingredients as appropriate to the desired preparation. It will be appreciated that the form and character of the pharmaceutically acceptable character or diluent is dictated by the amount of active ingredient with which it is to be combined, the route of administration and other well-known variables. The carrier(s) must be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not deleterious to the recipient thereof. The pharmaceutical carrier employed may be, for example, either a solid or liquid. Exemplary of solid carriers are iactose, terra alba, sucrose, talc, gelatin, agar, pectin, acacia, magnesium stearate, stearic acid and the like. Exemplary of liquid carriers are phosphate buffered saline solution, syrup, oil such as peanut oil and olive oil, water, emulsions, various types of wetting agents, sterile solutions and the like. Similarly, the carrier or diluent may include time delay

material well known to the art, such as glyceryl mono-stearate or glyceryl distearate alone or with a wax.

The dosage regimen will be determined by the attending physician and other clinical factors; preferably in accordance with any one of the above described methods. As is well known in the medical arts, dosages for any one patient depends upon many factors, including the patient's size, body surface area, age, the particular compound to be administered, sex, time and route of administration, general health, and other drugs being administered concurrently. Progress can be monitored by periodic assessment.

Furthermore, the use of pharmaceutical compositions which comprise antisenseoligonucleotides which specifically hybridize to RNA encoding mutated versions of the polynucleotitde or gene according to the invention or which comprise antibodies specifically recognizing a mutated polypeptide of the invention but not or not substantially the functional wild-type form is conceivable in cases in which the concentration of the mutated form in the cells should be reduced.

Thanks to the present invention the particular drug selection, dosage regimen and corresponding patients to be treated can be determined in accordance with the present invention. The dosing recommendations will be indicated in product labeling by allowing the prescriber to anticipate dose adjustments depending on the considered patient group, with information that avoids prescribing the wrong drug to the wrong patients at the wrong dose.

In another embodiment the present invention relates to the use of the polynucleotide, a polynucleotide comprising SEQ ID No: 104, a polynucleotide encoding a polypeptide comprising SEQ ID No: 145, the gene, the vector or the polypeptide of the invention, a polypeptide comprising SEQ ID No: 145 or the antibody of the invention for the preparation of a diagnostic composition for diagnosing a disease.

A gene encoding a functional and expressible polypeptide of the invention can be introduced into the cells which in turn produce the protein of interest. Gene therapy, which is based on introducing therapeutic genes into cells by *ex-vivo* or *in-vivo* techniques is one of the most important applications of gene transfer. Suitable vectors and methods for *in-vitro* or *in-vivo* gene therapy are described in the literature and are known to the person skilled in the art; see, e.g., Giordano, Nature Medicine 2 (1996), 534-539; Schaper, Circ. Res. 79 (1996), 911-919; Anderson, Science 256 (1992), 808-813; Isner, Lancet 348 (1996), 370-374; Muhlhauser, Circ. Res. 77 (1995), 1077-1086; Wang, Nature Medicine 2

(1996), 714-716; WO94/29469; WO 97/00957 or Schaper, Current Opinion in Biotechnology 7 (1996), 635-640, and references cited therein. The gene may be designed for direct introduction or for introduction via liposomes, or viral vectors (e.g. adenoviral, retroviral) into the cell. Preferably, said cell is a germ line cell, embryonic cell, or egg cell or derived therefrom, most preferably said cell is a stem cell. As is evident from the above, it is preferred that in the use of the invention the nucleic acid sequence is operatively linked to regulatory elements allowing for the expression and/or targeting of the polypeptides of the invention to specific cells. Suitable gene delivery systems that can be employed in accordance with the invention may include liposomes, receptor-mediated delivery systems, naked DNA, and viral vectors such as herpes viruses, retroviruses, adenoviruses, and adeno-associated viruses, among others. Delivery of nucleic acids to a specific site in the body for gene therapy may also be accomplished using a biolistic delivery system, such as that described by Williams (Proc. Natl. Acad. Sci. USA 88 (1991), 2726-2729). Standard methods for transfecting cells with recombinant DNA are well known to those skilled in the art of molecular biology, see, e.g., WO 94/29469; see also supra. Gene therapy may be carried out by directly administering the recombinant DNA molecule or vector of the invention to a patient or by transfecting cells with the polynucleotide or vector of the invention ex vivo and infusing the transfected cells into the patient.

A polynucleotide comprising SEQ ID No: 104 and a polypeptide comprising SEQ ID No: 145 have already been described in Jounaidi *et al.* (Jounaidi, Biochem Biophys Res Commun 221 (1996), 466-70). However, Jounaidi *et al.* have merely disclosed the respective amino acid and nucleotide sequences without making any suggestion towards the pharmaceutic and diagnostic value of said polynucleotide or polypeptide, in particular for those disorders and diseases referred to infra.

In a further embodiment the present invention relates to the use of the polynucleotide, a polynucleotide comprising SEQ ID No: 104, a polynucleotide encoding a polypeptide comprising SEQ ID No: 145, the gene, the vector, the polypeptide of the invention, a polypeptide comprising SEQ ID No.: 145 or the antibody of the invention for the preparation of a pharmaceutical composition for treating a disease.

In another embodiment the present invention encompasses the use of a polynucleotide selected from the group consisting of:

(a) a polynucleotide having the nucleic acid sequence of SEQ ID NO: 082, 088, 104, 112, 126, 131, or 140;

(b) a polynucleotide encoding a polypeptide having the amino acid sequence of SEQ ID No: 127, 132, 141 or 145;

- (c) a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said polynucleotide is having at least one additional nucleotide at a position corresponding to position 3709/3710 or 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614) or a nucleotide exchange at a position corresponding to position 7303 or 27289 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614);
- a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said (d) polynucleotide is having an additional G nucleotide at a position corresponding to position 3709/3710 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), an additional T nucleotide at a position corresponding to position 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), or an A at a position corresponding to position 7303 or 27289 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614);

for the preparation of a diagnostic composition for diagnosing a disease in a subject having a genome comprising a variant allele of the CYP3A5 gene, wherein said allele is having an A at a position corresponding to position 6986 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614).

The definitions of the terms referred to in this specification apply mutatis mutandis to the aforementioned use.

The term "subject" inter alia refers to animals. Preferably, said animals belong to the animal species referred to above. Moreover, the term "subject" encompasses humans. The humans in accordance with the use of the present invention are selected from all existing ethnical groups and subgroups, e.g. Caucasians, African Americans or Asians. However, particular well suited for the use of the invention are African Americans for which it could be demonstrated that diagnosing a disease or a prevalence for a disease based on monitoring the presence or absence of an CYP3A5 allele having at a position corresponding to position 6986 of the CYP3A5 gene (Accession No. AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614) an A results in a false positive prediction for CYP3A5 expression in a considerable number of subjects. This allele of CYP3A5 has been described in detail in Kuehl, 2001, Nature Genetics 27: 383-391 as CYP3A5*1 allele. The CYP3A5*1 allele is characterized by the presence of an A at position 22893 of the CYP3A5 nucleic acid sequence referred to in Kuehl, loc.cit. The alielic frequency of said allele is particularly high in African Americans although it is also present in other ethnical groups or subgroups. However, it was found in accordance with the present invention that the CYP3A5 expression of subjects for which a false positive result was obtained in diagnostic studies based on the CYP3A5*1 allele could be correctly predicted by further diagnosing the presence or absence of a polynucleotide as defined under (a) to (d) in accordance with the use of the present invention. For example, a polynucleotide having an additional nucleotide at a position corresponding to position 27131/27132 of the CYP3A5 gene as defined supra has been found in accordance with this invention to be present in approximately 10% of the African Americans resulting in a frameshift mutation in exon 11. The present invention provides means and methods to distinguish between the haplotype resulting in improved expression of CYP3A5 comprising the polymorphism(s) of the CYP3A5*1 allele and the haplotype resulting in decreased expression, wherein said haplotype as set forth above comprises in addition to the polymorphism(s) of the CYP3A5*1 allele co-segregating polymorphisms comprised by a polynucleotide referred to under (a) to (d) supra. Thus, based on the aforementioned use of the present invention a reliable diagnosis of the CYP3A5 activity of a subject is achieved.

The invention also relates to the use of a polynucleotide comprising a polynucleotide having an A at a position corresponding to position 14690 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614) for the preparation of a diagnostic composition for diagnosing a disease in a subject having a genome comprising a variant allele of the CYP3A5 gene, wherein said allele is having an A at a position corresponding to position 6986 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614).

The definitions of the terms referred to in this specification apply mutatis mutandis to the aforementioned use.

In accordance with the present invention it could also be shown that the polymorphism(s) constituting the CYP3A5*1 and the CYP3A5*6 allele as described in Kuehl, loc.cit., co-segregate in a considerable number of subjects and thereby constitute another haplotype resulting in decreased CYP3A5 expression. It has been shown that the CYP3A5*6 allele results in inappropriate splicing of exon 7 of CYP3A5, a frameshift and a premature termination at position 184 of the CYP3A5 protein. Consequently, false positive result as regards the expression level of CYP3A5 in a subject can be obtained in diagnostic studies based on the CYP3A5*1 allele. Said false positive results, however, can be avoided according to the use of this invention by further diagnosing the presence or absence of the polymorphism(s) of the CYP3A5*6 allele. Thus, based on the aforementioned use of the present invention a reliable diagnosis of the CYP3A5 activity of a subject is achieved.

In light with the foregoing, in a preferred embodiment of the aforementioned use said subject is an African American.

As has been discussed above, the number of subjects which are diagnosed false positive is due to the high allelic frequency of CYP3A5 alleles such as those comprising a polynucleotide as defined under (a) to (d) above resulting in a frame shift mutation or those comprised by the CYP3A5*6 allele. Said allelic frequency is particularly high within the group of African Americans. In accordance with the present invention it has been found that the CYP3A5*6 allele is present in about 13.3% of the African Americans. Thus within this ethnic group the problems emerging from a wrong prediction of CYP3A5 expression are more severe than for other ethnical groups.

In a more preferred embodiment of the use of the present invention said disease is cancer or diseases including cardiovascular diseases, diabetes and AIDS.

Finally, the present invention relates to a diagnostic kit for detection of a polymorphism comprising the polynucleotide, the gene, the vector, the polypeptide, the antibody, the host cell, the transgenic non-human animal or the solid support of the invention.

The kit of the invention may contain further ingredients such as selection markers and components for selective media suitable for the generation of transgenic cells and animals. The kit of the invention can be used for carrying out a method of the invention and could be, inter alia, employed in a variety of applications, e.g., in the diagnostic field or as research tool. The parts of the kit of the invention can be packaged individually in vials or in combination in suitable containers or multicontainer units. Manufacture of the kit follows preferably standard procedures which are known to the person skilled in the art. The kit may be used for methods for detecting expression of a mutant form of the polypeptides, genes or polynucleotides in accordance with any one of the above-described methods of example. immunoassay techniques invention, employing. ihe radioimmunoassay or enzymeimmunoassay or preferably nucleic acid hybridization and/or emplification techniques such as those described herein before and in the Examples as well as pharmacokinetic studies when using non-human transgenic animals of the invention.

The figures illustrate the invention.

Figure 1: A. Western blot analysis of CYP3A5 protein expression in microsomes prepared from 6 LE (low expressing) and 6 HE (high expressing) Caucasian livers. **B**. The relative contributions of CYP3A5 and CYP3A4 to the combined CYP3A5/CYP3A4 protein pool in 17 HE livers as determined by Western blot.

Figure 2: Expression levels and the allelic source of *CYP3A5* transcripts in LE and HE Caucasian livers. *A*. TaqMan analysis of *CYP3A5* mRNA in 8 LE (low expressing, white bars) and 8 HE (high expressing, grey bars) liver samples. *B*. Sequences of a portion of the 3'-UTR of *CYP3A5* in samples heterozygous for variant ch-v-015 (Table 2A). Templates used for PCR were genomic DNA (left panel), cDNA from a LE liver (middle panel) and cDNA from an HE liver (right panel).

Figure 3: Allelic frequencies of *CYP3A5* genetic variants in Caucasians. **A.** in DNA samples derived from 8-168 LE individuals **B.** in DNA samples derived from 7-18 HE individuals. The bars at the bottom of the figure indicate schematically the localisation of the pseudoexons PS2 exon 1 and 2 and exons 1 – 13 of the CYP3A5 gene. The arrowhead marks the duplication boundary (Gellner, Pharmacogenetics 11 (2001), 111 – 121).

Figure 4: Genomic and peptide sequences: genomic DNA sequences containing the amplified regions in which polymorphisms were detected and polypeptide sequences with amino acid substitutions. Nucleotide sequences are listed in 5' - 3' orientation. Letters in lowercase indicate non-coding sequences, letters in uppercase indicate coding sequences. Primer regions are underlined. Variant sites are shown framed. Peptide sequences are shown in one letter code. || marks a site where a deletion has occurred. In Seq ID 198, the hybridzing site of the TaqMan® probe is shown in bold.

Figure 5: CYP3A5 cDNA insert region of the plasmid that was used as starting material for in vitro mutagenesis. Cloning sites are shown underlined. Modified 5' and 3' regions of the CYP3A5 cDNA are shown in lowercase letters. The 5' modification, a MALLLAVF amino acid sequence on protein level, has been introduced in order to increase expression in *E. coli* (Gillam, Arch Biochem Biophys 317 (1995), 374-84). The 3' modification, a His₆ tag on protein level, has been introduced in order to enhance subsequent purification. The unmodified part of the CYP3A5 insert was verified to be identical to the CYP3A5 cDNA corresponding to accession no. NM_000777.1 by sequencing, which is the underlying nucleotide sequence for NP_000768.1. Sites corresponding to ch-v-009 and ch-v-001 are shown framed.

The invention will now be described by reference to the following biological Examples which are merely illustrative and are not constructed as a limitation of the scope of the present invention.

Example 1: Isolation of genomic DNA from human blood, generation and purification of CYP3A5 gene fragments

Genomic DNA was isolated from blood or liver samples using Qiagen blood and tissue DNA isolation kits. Oligonucleotides used in the screen were designed based on the recently determined sequence and organisation of the human *CYP3A* locus (Gellner, Pharmacogenetics 11 (2001), 111 - 121). Primer sequences and PCR fragment lengths are given in Table 1A. Amplified fragments were processed through PCR purification columns (Qiagen) and sequenced on PE ABI 3700 DNA Analysers using the same primers as in PCR. The sequences were analysed for the presence of polymorphisms using the PHRED/PHRAP/POLYPHRED/CONSED software package (University of Washington, Seattle, WA, USA).

Total RNA was isolated from liver samples using the RNeasy kit (Qiagen) according to the manufacturers instructions except that an additional DNase I digestion was performed directly on the column. cDNA pools were generated from 1 μ g of total RNA using random hexamer primers and Superscript reverse transcriptase (Life Technologies). The cDNA used for one TaqMan assay was derived from 40 ng total RNA. *CYP3A5* mRNA expression levels were quantified by real time quantitative PCR using the ABI 7700

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Sequence Detection System (PE Biosystems). Oligonucleotides and probes were designed with the Primer Express (PE Biosystems) programme. Oligonucleotides used for the quantitative PCR were: forward 5'- TTG TTG GGA AAT GTT TTG TCC TAT C -3' (Seq ID: 237) and reverse 5'- ACA GGG AGT TGA CCT TCA TAC GTT -3' (Seq ID: 238). The TaqMan probe (5'- TCA GGG TCT CTG GAA ATT TGA CAC AGA GTG CTA-3'; Seq ID: 239) was labelled with the 5' reporter dye 6-carboxy-fluorescein (FAM) and the 3' quencher 6-carboxy-tetramethylrhodamin (TAMRA). The experiments were performed according to a standard protocol developed by PE Biosystems. The specificity of the assay for *CYP3A5* was determined using equal amounts of *CYP3A4*, *CYP3A5*, *CYP3A7* and *CYP3A43* cDNA species expressed *in vitro*. The specificity of the probe was 10⁴ times higher for *CYP3A5* than for *CYP3A7* cDNA whereas *CYP3A4* and *CYP3A43* cDNAs were not detectable at all. The linear range of the *CYP3A5* assay was determined to be between 10 and 10⁶ target molecules. *CYP3A5* expression levels were normalised using the expression of 18S mRNA species determined with pre-developed TaqMan assays (PE Biosystems).

Example 2: Determination of genetic variations within the CYP3A5 locus

Sequence diversity within the CYP3A5 locus was determined by PCR amplification from genomic DNA (fragment size: 264 - 997 bp) and sequencing each PCR-product of 19 -217 samples of Caucasian origin, 36 - 45 samples of African American origin, 34 - 47 samples of Chinese origin, 41 - 50 samples of Japanese origin, and 31 - 47 samples of Korean origin. The PCR fragments encompass the entire protein-coding region of CYP3A5, a portion of the 3'-UTR, the entire 5'-UTR as well as 6203 bp sequence between the CYP3A5 transcriptional start site and a L1_5'UTR_ORF repeat located upstream of the gene (Fig. 3). In addition, we genotyped two linked single nucleotide polymorphisms (SNPs, ch-v-020, ch-v-021, Table 2A-E) located in a sequence originally described as CYP3A5 promoter that were recently reported to co-segregate with increased CYP3A5 protein expression (Paulussen, Pharmacogenetics 10 (2000), 415-24). The results also indicate co-segregation of both variants. Using the recently determined sequence of the entire CYP3A locus (Gellner, Pharmacogenetics 11 (2001), 111 - 121), we place these variants approximately 20 kb upstream of the first exon of CYP3A5, in a sequence 5' adjacent to a CYP3A pseudogene (PS2 in Fig. 3). Furthermore, we additionally genotyped a single nucleotide polymorphism located in intron 3 of the CYP3A5 gene (ch-v-048;

Kuehl, 2001, Nature Genetics 27: 383-391) by TaqMan® assay using the primers and probes listed in Table 1B. The analysis were performed on a Sequence Detection System (PE Biosystems).

A total of 29 variants including the two linked SNPs described by Paulussen et al. (Paulussen, Pharmacogenetics 10 (2000), 415-24) were detected in the screen of Caucasian samples and their allelic frequencies were estimated to be between 0.3 % and 11.9 % (Table 2A). 6 variants are located within the 6 kb sequence upstream of the transcriptional start site of *CYP3A5*. 14 variants are located in introns, or in the 5'-UTR or 3'-UTR, whereas 4 have been found in the protein-coding sequence. Among the latter ones, three variants result in amino acid substitutions and one in a premature termination of the CYP3A5 protein (Table 2A). The g.7303C>A variant (ch-v-009, Table 2A) results in a S100Y amino acid exchange in exon 4. The g.3705C>T variant (ch-v-005) leads to a H30Y amino acid exchange in exon 2. Cloning and sequencing revealed a physical linkage of this variant to the g.3709-3710insG variant (ch-v-006). The latter variant results in a shift of the open reading frame leading to a truncation of the protein sequence at position 34 (K34.). The T398N variant (ch-v-001, Table 2A), originally described by Jounaidi (Jounaidi, Biochem Biophys Res Commun 221 (1996), 466-70), was found in 3 out of 80 individuals tested.

Neither of the four protein altering variants found in Caucasians have been found in the African-Americans, Chinese, Japanese or Korean samples (Table 2A-E). However, among others we have found 4 new variants in these samples that result in an altered CYP3A5 amino acid sequence (ch-v-017, ch-v-043, ch-v-045, ch-v-068, Table 2B-E). The g.27131-27132insT (ch-v-017) variant in exon 11 (ch-v-017, Table 2B, 2D) has been found in 9 out of 45 African-American samples and in one out of 50 Japanese samples. The variant results in a shift of the open reading frame which leads to a truncation of the protein sequence at position 348 (D348.). Variants ch-v-043, ch-v-045 and ch-v-068 lead to amino acid exchanges.

Example 3: Identification of genetic determinants of CYPA5 protein expression

In the following, the frequencies of Caucasian CYP3A5 gene variants have been analyzed as a function of CYP3A5 protein expression. For this purpose, allelic frequencies of

variants shown in Table 2A were calculated separately for HE and LE livers (Fig. 3). The frequencies of 9 variants (ch-v-020, ch-v-021, ch-v-026, ch-v-034, ch-v-007, ch-v-008, ch-v-011, ch-v-014 and ch-v-015) were significantly increased in HE livers (all $\chi^2 > 13.3$, df = 1, p < 0.01, Bonferroni corrected). Except one, all tested HE livers (17/18, 94 %) were heterozygous for three variants (ch-v-021, ch-v-026 and ch-v-015). 16 of those samples were heterozygous for ch-v-020 as well. One HE sample could not be genotyped for this variant. In contrast, LE livers were either wildtype (155/168, 92.3 %), heterozygous for variants ch-v-021 and ch-v-26 (9/168, 5.4 %) or heterozygous for the variant ch-v-015 (4/168, 2.4 %) only. However, in LE livers all three variants never occurred simultaneously (Table 3). These results defined either of the three variants as a useful but imperfect marker of increased CYP3A5 expression. The variants ch-v-034, ch-v-008, ch-v-011 and ch-v-14 only occurred in a subset of the samples heterozygous for the above three variants (ch-v-021, ch-v-026 and ch-v-015).

The distribution of variants ch-v-021, ch-v-026 and ch-v-015 in the samples screened strongly suggest that they constitute a haplotype. In the following, the hypotheses whether these three variants recombine independently or not has been tested. Assuming their independent inheritance, the expected 3-loci-genotype frequencies for all combinations of variants and compared them with the observed frequencies have been calculated. The difference is highly significant ($\chi^2 = 93.6$; classes 'all wildtype', 'single variant hetero- or homozygous', 'two or three variants hetero- or homozygous'; df = 1; p << 0.001). There were more individuals with two or three of the variants than expected and less individuals with only one of the variants. This result suggests linkage among the three variants. The degree of linkage with the linkage disequilibrium parameter D for the three pairs of variants was estimated. Using maximum likelihood estimates for haplotype frequencies, D was calculated to be 0.041 for the variant pairs ch-v-021/ch-v-015 and ch-v-026/ch-v-015, which is 80 % of its theoretical maximum, and 0.065 for variants ch-v-021 and ch-v-026 which corresponds to 100 % of its theoretical maximum.

The probability that individuals showing the respective variant genotype are HE (positive predictive value) is estimated to be 65 % for variants ch-v-021 and ch-v-026, respectively, and 81 % for the ch-v-015 variant. For the combination of all three variants the positive predictive value is 100 % in our sample set. However, assuming that these variants need to be located in *cis* for increased protein expression, it is clear that there is some probability for individuals showing all three variants to be LE. The results show that at least the allele ch-v-021/ch-v-026 and the allele ch-v-015 actually exist (see genotype 2 and 3,

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Table 3) and therefore the existence of a genotype with a combination of these two alleles has to be postulated. The maximum likelihood estimate for the frequency of these 3-fold heterozygotes having not all three variants in *cis* is 0.05 % of all samples screened or 0.61 % of samples hetero- or homozygote for all three variants. In other words, of 100 Caucasians screened statistically about 9 of them will be hetero- or homozygous for all three variants and about 0.05 of these will have not all three variants in *cis*. Therefore, it can be expected that the positive predictive power of the 3-variant genotype to be about 99.95 %. Of course, the same values would be achieved for a combination of only two variants, either ch-v-021/ch-v-015 or ch-v-26/ch-v-15.

In a single HE liver none of the above 9 variants that were found in the other 17 HE samples could be detected. A closer examination of variants found in this sample revealed a variant within intron 4 (ch-v-018) and one within intron 5 (ch-v-019), respectively. These variants were unique to this sample, since they were not found in any other of the samples screened. Neither were they found in any of the other ethnic groups screened. It remains to be shown whether these variants are themselves causative for transcriptional activation or whether they are linked to another, so far undetected variant.

Example 4: Determination of CYP3A5 protein expression

Protein expression of CYP3A4 and CYP3A5 in Caucasian liver samples was determined by Western blotting using CYP3A4- and CYP3A5-specific antibodies (Gentest). Liver microsomes were prepared as previously described (Zanger, Biochemistry 27 (1988), 5447-54). To obtain total protein homogenate, powdered liver tissue was homogenised in 0.1 M Tris-Cl pH 7.4, 1 mM EDTA, 1 mM Pefa Bloc SC, 1 µg/ml leupeptin, 1 µg/ml pepstatin with a Potter Elvehjem homogenisator (glass/Teflon) for 2 min at 1000 rpm. Homogenates were then sonified with a Bandelin Sonoplus HD 200 and stored at –80°C. For Western blotting, 12.5 µg microsomal protein homogenate or 40 µg total protein homogenate were separated in a 10 % SDS-polyacrylamide gel. Electrophoretic transfer onto PVDF membranes was carried out in a TankBlot Cell (BioRad) for 1.5 hours at constant voltage (100 V) and at 10 °C. Following the transfer, the membranes were incubated for 60 min in 5 % milk, TBS, 0.1 % Tween 20 to reduce the unspecific antibody binding. Incubations with either primary antibody (Gentest, dilution 1:500) were performed in 1 % milk, TBS, 0.1 % Tween 20 for 60 min, those with the secondary antibody (anti-

rabbit IgG-POD Fab-fragments, Dianova, dilution 1:10000 in the same solution for 30 min. CYP3A4 or CYP3A5 protein bands were detected with Supersignal Dura (Pierce) and a digital CCD-camera (LAS-1000, Fuji). Signal quantification was performed with AIDA (Raytest). Protein expression levels were calculated based on calibration curves obtained with microsomes expressing recombinant CYP3A4 and CYP3A5 proteins (Gentest).

Homogenates or microsomal fractions were prepared from 186 human livers and investigated by Western blotting using a CYP3A5-specific antibody. CYP3A5 protein was detected in all samples analysed and its expression showed a clear bimodality (Fig. 1A). 168 livers (~ 90 %), further referred to as LE (low-expressing), showed expression close to or below the lower limit of quantification (LLOQ) of the assay (0.3 pmol/mg homogenate protein and 1.0 pmol/mg microsomal protein). Eighteen samples (~ 10 %), further referred to as HE (high-expressing), exhibited much higher CYP3A5 expression levels. The expression was in the range between 1.6 and 2.9 pmol/mg homogenate protein (2.3 \pm 0.5; n = 6) and between 3.9 and 15.5 pmol/mg microsomal protein (9.7 \pm 4.1; n = 12). Taking the LLOQ of the assay as the expression level of CYP3A5 in LE livers, HE livers express on average 8 to 10 times more CYP3A5 protein than LE livers.

In the following, the contribution of CYP3A5 to the combined CYP3A5 and CYP3A4 protein expression in HE livers was investigated. CYP3A4 expression in these livers was between 0.9 and 82.6 pmol/mg homogenate protein (n = 6) and between 4.5 and 295 pmol/mg microsomal protein (n = 11). The levels and the range of CYP3A4 variability in HE livers were similar to those in LE livers (not shown). Fig. 1B shows the share of CYP3A5 in the combined CYP3A5 and CYP3A4 protein pool in 17 HE livers. CYP3A5 contribution varies between 3 % and 74 %. In an average HE liver, the share of CYP3A5 in the combined pool of both proteins is 24 %. Taking the LLOQ of the CYP3A5 assay as the actual expression level of the protein in LE livers, the corresponding value in these livers is approximately 1.6 %.

Example 5: Determination of CYP3A5 mRNA expression

The expression of CYP3A5 mRNA in 8 Caucasian HE and 8 LE livers was investigated using a CYP3A5-specific TaqMan probe. As illustrated in Fig. 2A, the distribution of CYP3A5 mRNA levels exhibited a bimodality which was in complete agreement with that

observed in the expression of CYP3A5 protein (Fisher's exact test, p = << 0.001). The number of 3A5 transcripts per ng of total RNA in HE livers (n = 8) was on average 8.5 times higher than those in LE livers (n = 8).

In the following, the allelic origin of *CYP3A5* transcripts in HE and LE livers was investigated. To this end, by PCR a portion of the 3'-UTR (untranslated region) of the gene was amplified and sequenced using genomic or cDNA samples as templates which were heterozygous for a T>C variant located in this region (variant ch-v-015 in Table 2A;). As expected, both alleles are represented in the sequence using genomic DNAs as template (Fig. 2B). Both alleles were also equally represented in a sequence of PCR-amplified cDNA from a LE (homozygous wildtype for ch-v-021 and ch-v-26, heterozygous for ch-v-015) liver. In contrast, only the C allele was found in the same portion of *CYP3A5* 3'-UTR cDNA from a HE liver. This indicates an overrepresentation of transcripts derived from the chromosome harbouring the C allele in the total pool of *CYP3A5* transcripts in HE livers.

Example 6: In vitro mutagenesis and expression of recombinant CYP3A5 proteins

Five polymorphisms in Caucasians have been detected that lead to changes in the protein sequence. Two of them, ch-v-006 and ch-v-017, lead to a truncation of the protein and therefore are unlikely to code for a functional protein. As ch-v-005 has only been found physically linked to ch-v-006, the resulting protein variant is not likely to be functional as well. To determine the effect of the protein variants ch-v-009 and ch-v-001 these variants were analysed in a heterologous bacterial expression system.

A modified CYP3A5 cDNA in the prokaryotic expression vector pKK233-2 (Pharmacia) was used as starting material for in vitro mutagenesis (Fig. 5). Variants ch-v-009 and ch-v-001, respectively, were introduced into the plasmid by in vitro mutagenesis using the QuikChange mutagenesis kit (Stratagene). The successful introduction of the mutations and the absence of other, undesired mutations was confirmed by sequencing. The original plasmid as well as the two mutagenised plasmids were used to transform *E. coli* TOPP3 cells, a strain in which optimal expression of CYP3A proteins has previously been obtained. A total of 8 separate colonies of each mutant plasmid were chosen for expression studies. The bacteria were grown and induced as described in Eiselt *et al.* (Eiselt, Pharmacogenetics 11 (2001), 447-58.). Expression was analysed 48 h and 72 h after induction with IPTG/δ-ALA. Cells were harvested as described in Domanski *et al.*

(Domanski, Arch Biochem Biophys 350 (1998), 223-32). The final P450 content was measured by reduced carbon monoxide (CO) difference spectra (Omura, J. Biol. Chem. 239 (1964), 2370-2378).

Whereas 30 to 50 nmol solubilised CYP3A5 could be recovered per litre culture of the non-mutagenised CYP3A5, expression in the two CYP3A5 variants S100Y and T398N was determined to be lower than 3 nmol P450 protein per litre culture. In many instances, the "P450" peak was shifted to 454 - 458 nm rather than the typical 448 - 450 nm peak expected. The low level of expression in mutagenised colonies made any attempts at protein purification futile. Previous experiments suggest that expression levels as low as those demonstrated by these CYP3A5 variants can not be significantly improved by utilising other bacterial strains or adjusting growth temperature. Therefore, the results strongly suggest that the CYP3A5 protein variants comprising the S100Y or the T398N substitutions are unstable in an *E. coli* expression system and that the variants comprising ch-v-009 or ch-v-001 may not code for functional proteins. The result of negative expression for variant ch-v-001 (T398N) is in agreement with the study in which this polymorphism was initially detected in two of five CYP3A5 deficient individuals (Jounaidi, Biochem Biophys Res Commun 221 (1996), 466-70).

Example 7: Prediction of expected drug metabolism by CYP3A5 genotypes

The CYP3A5 protein degrades many drugs by oxidation so that they are not therapeutically active anymore. Therefore, drugs that are CYP3A5 substrates might not reach therapeutically active plasma concentrations for an adequate time span in patients with enhanced CYP3A5 activity. In these patients these drugs have to be dosed higher. On the other side, in patients with reduced CYP3A5 activity, these drugs have to be administered at lower dosage in order to avoid toxic drug levels. Table 4 gives an assignment for CYP3A5 genotypes and recommended dosages.

In cases in which CYP3A5 metabolism leads to the formation of pharmacologically active substance, enhanced enzyme activity has to be counteracted by reduced dosage whereas reduced CYP3A5 activity has to be met by increased dosage.

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Table 1A: Primers used to screen for polymorphisms within the *CYP3A5* upstream regions and the *CYP3A5* gene.

			Primer	Position (nt)	(bp)
Ref.	ID	Name	Sequence (5'-3')	Position (nt)	(nh)
	001	694	ACAGGCACAGAACCCACAAG	145448-145468 ¹	630
chzk	002	711	ATCGCCACTTGCCTTCTTC	146077-146059	030
	003	794	CCCTGCTTCGGCTTGTGCA	159915-159933	575
chzl	004	750	CACAGCCTGCTTTATTTGTCATGA	160489-160466	5/5
	005	751	GATCCTTGGTAGGACAAGCCT	160351-160371	844
chzj	006	754	CAAGCACTGATTTGGTCACTTCCT	161194-161171	044
	007	819	GGGATGGGACCGTAAGTGGAAC	160951-160972 1	618
chzb	008	820	TAATCACATTGGAGTTCTGACAAATG	161568-161543 ¹	010
	009	736	AAAAACCTCTTACAAAAGTATCATCGGATA	161419-161448 ¹	910
chzi	010	737	CCTACTAGGTCTCTGACTTGGAACCAT	162328-162302 ¹	910
	011	784	GCCGAGACGCACCATTACACT	161876-161896 ¹	607
chzh	012	785	CACCCATCCCTTCCCACTCAT	162512-162492	637
	013	740	TGATGGTTCCAAGTCAGAGACCTAGTAG	162300-162327	007
chzg	014	741	AATTGTAGACATCTTTCTCTTAAGTTAATTCCCAG	163296-163262	997
	015	786	TCTGCATGCCAACAGTGAACAATCT	163182-163206 ¹	004
chzf	016	789	GGCACGCACCAGCATGTCC	164005-163987	824
	017	790	CTGGCTGAGTGCCGTGGCT	163845-163863	CO1
chze	018	791	TGAGCGCTTCATGTATTCTGGCTAT	164435-164411	591
	019	824	AAATATTTTCAAAGTCACACTCTGACAACAG	164376-164406	0.7.7
chza	020	822	TAACAGGATCTCATGCTTTTTTCATGGCT	164992-164964	617
	020	747	CACTCCAATATTCACAATAGCCACTATTCA	164843-164872	200
chzd	022	748	ACTCCTACGTATCCTTCCAAGCCC	165768-165745	926
	023	728	GCTAAGGGAAACAGGCATAGAAACTTAC	165586-165613	
chzc	023	729	GGAGCTTCCCTGCCCTGC	166142-166125 1	557
	025	323	TCCTTCTCCAGCACATAAATC	166076-166096	404
chzy	026	325	AAATTAGAAGGTGGATGGGAG	166499-166479	424
	027	335	GAGTAACTCACCAGCCCTCTG	·169838-169858 1	004
chzx	028	336	AAACCTCAGAACTCCCTCCCA	170101-170081	264
	020	338	GACATCTCTGAATAGCTTCCTTC	171392-171414	400
chzw	030	341	GCACATAGTTTATAACGGCAA	171793-171773	402
	030	346	AGAACCTAAGGTTGCTGTGTC	173303-173325	201
chzv	032	348	TGCAAGATGTTACCACTGGGC	173696-173676	394
		354	CGCCCACATACACTCAGAA	31376-31395 ²	400
chzu	033	357	AGACCATTTTTAGGAAGCTCG	31801-31781 ²	426
	035		CAAGGGTAGTCCACTGAGTTC	31760-31781 ²	400
chzt	036		CTCTTTGGAGTTGCAGCG	32162-32145 ²	403
	037		AGGTGAGTCTAACTCAGCTTG	33081-33101 ²	570
chzs			GACAGCTAAAGTGTGTGAGGG	33658-33638 ²	578
	038		AATGGGTTCCAGTTGAGAATC	34411-34431 ²	470
chzr	039		ATTGTTGTGCCTGATTTCAAG	34880-34860 ²	470
			AGAAGCCATAGGGAGGTTG	35627-35645 ²	
chzq	041		GACTGTCCTCCAAGCATTCT	36049-36030 ²	423
			GATGCCATGATGAGGAGTGTG	37724-37744 ²	
chzp	043		ACCAGGGCCAGCAATATTG	38349-38331 ²	626
<u> </u>	044		AAATACTTCACGAATACTATGATCA	45711-45735 ²	
chzm	045		CAGGGACATAATTGATTATCTTTG	46305-46282 ²	595
ļ 	046		TACTGGTTGGGAGGTGGAG	48290-48308 ²	1
chzo	047		CATGATGTTCTTAATGCTACAGG	48745-48723 ²	456
	048		GAAGAGTTCAAGATACATGGTGTTA	50088-50112 ²	
chzn	049		TGCACAACACTCTACACAGACTC	50503-50481 2	416
 -	050	420	TGCACAACACTCTACACAGACTC	- 1- ODords 00-	ـــــ

Ref.: Reference sequence. The positions of primers refer to GenBank sequences with accession numbers (1) AF280107.1 and (2) AC005020.2.

Table 1B: Primers (Seq IDs 202 and 203) and probes (Seq IDs 204 and 205) used determine the nucleotide status at the polymorphic site ch-v-048 (g.6986G>A) by

TagMan assay.

, agin			Primer	Position (nt)	(bp)
Ref.	ID	Name	Sequence (5'-3')	rosidon (iit)	(44)
	202	TQPi_ch-v-048_F	GCTCTACTGTCATTTCTAACCATAAT CTCTTTA	173152-173184	
	204	TQPo_ch-v- 048_Al1_G_VIC	VIC-TGTCTTTCAGTATCTCTT- MGB-DQ	173196-173213	99
chyu	205	TQPo_ch-v- 048_Al2_A_FAM	FAM-TGTCTTTCAATATCTCTTC- MGB-DQ	173196-173214	
	203	TQPi_ch-v-048_R	GCTTCATATGATGAAGGGTAATGTGG T	173250-173224	

Ref.: Reference sequence. The positions of the primers refer to the GenBank sequence with accession number AF280107.1. Probes are labelled with a fluorenscent dye at the 5' end and labelled with a dark quencher (DQ) and using a minor groove binder (MGB).

Table 2A: CYP3A5 polymorphisms detected in samples of Caucasian origin.

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Variant	Reference	Variant po	position on	Sedi	Sequence context	Genetic	Predicted	Car	Caucasian
Ω	sednence	Reference sequence	gDNA	Seq	Reference seq. Variant seq.	element	effect	Z	Variant allele frequency (%)
ch-v-020	chzk	254T>G	g20619T>G	051 052	TGCGCITGCAA	5' of PS2		211	9.9
ch-v-031	chzk	318G>A	g20555G>A	053 054	GCATGGGTAAA	5' of PS2		189	0.3
ch-v-032	chzk	544G>A	g20329G>A	055 056	GGGGTGTGTGC	5' of PS2	,	186	0.3
ch-v-033	chzk	550G>A	g20323G>A	057 058	TGTGCGATTCT	5' of PS2		186	0.3
ch-v-021	chzk	582A>G	g20291A>G	059 060	GCCCCACCTCC	5' of PS2		215	6.7
ch-v-026	chzl	229A>G	g6177A>G	061 062	CTCACACTGGG	5' of Exon 1		208	7.0
ch-v-027	chzi	566G>A	g4336G>A	063 064	GAGACGCACCA	5' of Exon 1		19	2.7
ch-v-028	yzyo	601G>A	g3844G>A	065 066	TGTGTGTGGGA	5' of Exon 1		20	10.0
ch-v-029	gzyo	464T>C	g3557T>C	067 068	ATCCATGTATA	5' of Exon 1		20	2.5
ch-v-034	chza	328T>C	g1617T>C	069 070	CATCTTACCCC	5° of Exon 1		66	2.2
ch-v-030	chzd	683T>A	g795T>A	071 072	TCTATTGCTAT	5' of Exon 1		20	5.0
ch-v-002	chzy	159G>A.	g86G>A	073 074	GGCAGGGAAGC	Exon 1 (5' UTR)		106	0.5
ch-v-003	chzy	171C>T	g74C>T	075 076	CCAGGCAAACA	Exon 1 (5' UTR)		106	4.3
ch-v-004	chzy	418-420delGAG	g.174-176delGAG	077 078	TCAAGGAGAAG	Intron 1		106	0.5
ch-v-005	chzx	187C>T	g:3705C>T	079 080	GTACACATGGA	Exon 2	Н30У	104	1.4
ch-v-006	chzx	191-192insG	g.3709-3710irisG	081 082	CATGG-ACTTT	Exon 2	(K34:) 1	104	1,4
ch-v-007	chzw	143C>T	g.5215C>T	083 084	GATAGCAGGCC	Intron 2		105	6.7

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ch-v-048	chyu	206G>A	g.6986G>A	147	1010101011	Intron 3	splice defect	217	4.6
ch-v-008	chzv	199C>A	g.7182C>A	085 086	AGAATCGGGCT	Intron 3		107	1.9
ch-v-009	chzv	320C>A	g.7303C>A	087	TTATICTGTCT	Exon 4	S100Y	107	0.5
ch-v-018	chzv	441- 444insCTAAAAAT	g.7424- 7427insCTAAAAAT	060 680	CAGG CCTAAAAATG	Intron 4		107	0.5
ch-v-016	chzt	145T>G	g.13077T>G	091 092	TCTTTTTTT	Intron 5		105	0.5
ch-v-019	- chzt	241T>C	g.13173T>C	093 094	GAGTCTGCACA	Intron 5		105	0.5
ch-v-010	chzq	132-133insGTC	g.16931- 16932insGTC	095 096	AGTCAAGA	Intron 8		95	0.5
ch-v-011	chzq	364G>T	g.17163G>T	260 260	AGGAAGTATTC	Intron 9		95	3.2
ch-v-012	chzp	269G>A	g.19165G>A	001	AGAGAGCTTCA	Intron 9		106	0.5
ch-v-013	chzm	167A>G	g.27050A>G	101	CTTCAATAGTA	Intron 10		80	11.9
ch-v-001	chzm	406C>A	g.27289C>A	103 104	TCCAACTTATG	Exon 11	. T398N	80	1.9
ch-v-014	chzm	643C>T	g.27526C>T	105 106	CGAAACTACAT	Intron 11		80	3.8
ch-v-015	chzn	351T>C	g.31611T>C	107	AAGGATTTCTA	Exon 13 (3' UTR)		197	5.6
All varies	nte word	All variants were detected in the betar	F 207 400000 04040 011000 1200	1 5 2 2 3	1			. 070	

All variants were detected in the heterozygous state except for 1 homozygous individual for variant ch-v-013. 1 variant results in a frame shift which ultimately leads to a premature termination.

Table 2B: CYP3A5 polymorphisms detected in samples of African-American origin.	Variant position on Sequence context Genetic Predicted N Variant allele	Reference sequence gDNA ID variant seq. element effect ' frequency (%)	230T>C g20643T>C 148 TTTAATAGAAG 5' of PS2 42 3.6	254T>G g20619T>G 051 TGGGCTTGCAA 5' of PS2 41 69.5	506C>T g20367C>T 150 ATTCCCCATAG 5' of PS2 44 1.1	514T>C g20359T>C 152 TAGAATATGAA 5' of PS2 44 1.1	582A>G g20291A>G 059 GCCCAACCTCC 5' of PS2 45 66.7	229A>G g6177A>G 061 CTCACACTGGG 5' of Exon 1 43 65.1	455T>G g3990T>G 154 GTAACTTATCC 5' of 44 2.3	TTCA	601G>A g3844G>A 065 TGTGTGTGGGA 5' of Exon 1 44 17.1	328T>C g1617T>C 069 CATCTTACCCC 5' of Exon 1 45 42.2	159G>A g86G>A 073 GGCAGGGAAGC Exon 1 45 1.1	171C>T g74C>T 075 CCA3GCAAACA Exon 1 45 1.1	143C>T 083 GATAGCAGGCC Intron 2 44 3.4	163G>A 158 TGGACGCAACT Intron 2 45 2.2		_
of African-Americ	Sequence conte				<u> </u>						-		-		-		-	
etected in samples	sition on	BDNA	g20643T>C	g20619T>G	g20367C>T	g20359T>C	g20291A>G	g6177A>G	g3990T>G	g3868G>A	g3844G>A	g1617T>C	g86G>A	g74C>T	g.5215C>T	g.5235G>A	g.5516T>A	
A5 polymorphisms de	Variant pos	Reference sequence	230T>C	254T>G	506C>T	514T>C	582A>G	229A>G	455T>G	577G>A	601G>A	328T>C	159G>A	171C>T	143C>T	163G>A	444T>A	
2B: CYP3,	Reference	sednence	chzk	chzk	chzk	chzk	chzk	chzl	chzh	chzh	chzh	chyz	chzy	chzy	chzw	chzw	chzw	
Table	Variant	۵	ch-v-037	ch-v-020	ch-v-038	ch-v-039	ch-v-021	ch-v-026	ch-v-051	ch-v-052	ch-v-028	ch-v-034	ch-v-002	ch-v-003	ch-v-007	ch-v-053	ch-v-054	

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7.0	1.1	1:	13.3	6.7	1.1	6.7	3.4	1.1	7.0	1.1	2.2	4.1	10.0	11.9	5.6	68.2
43	45	45	45	45	45	45	44	44	43	45	45	36	45	42	45	44
	1149T		splice defect	P218P									(D348.) ¹			
Intron 3	Exon 6	Intron 6	Exon 7	Exon 7	Intron 7	Intron 7	Intron 7	Intron 8	Intron 9	Intron 9	Intron 9	Intron 10	Exon 11	Intron 11	Intron 12	Exon 13 (3' UTR)
AGCTCCGTTGT	CATCATTGCCC	CAGICGCACTG	ACTAAGAAGTT	GATCCATTATT	CAATTCCATTG	TGTCAATCTAG	TTGTTTGTTT	AAATAAAGAAG	AGGAAGTATTC	TTTGCGTCATC	TTGACCTGATT	CTTCAATAGTA	CACCT-ACCTA	CGAAACTACAT	TATTGTAGATCC.	AAGGATTTCTA
109	162 163	164	1	168 169	170	172 173	174 175	176	260 260	178	180	101	111		182 183	107
g.7207C>T	g.13226T>C	g.13376G>A	g.14690G>A	g.14720A>G	g.14836C>T	g.14903A>G	g.15788T>C	g.16079A>C	g.17163G>T	g.19069G>A	g.19208C>T	g.27050A>G	g.27i31-27132insT	g.27526C>T	g.31499T>C	g-31611T>C
224C>T	294T>C	444G>A	437G>A	467A>G	583C>T	650A>G	205T>C	496A>C	364G>T	173G>A.	312C>T	167A>G	248-249insT	643C>T	239T>C	351T>C
chzv	chzt	chzt	chzs	chzs	chzs	chzs	chzr	chzr	chzq	chzp	chzp	chzm	chzm	chzm	chzn	chzn
ch-v-025	ch-v-043	ch-v-055	ch-v-050	ch-v-056	ch-v-057	ch-v-058	ch-v-059	ch-v-060	ch-v-011	ch-v-062	ch-v-063	ch-v-013	ch-v-017	ch-v-014	ch-v-044	ch-v-015

variant results in a frame shift which ultimately leads to premature termination.

Reference sequence Variant position on sequence context Sequence sequence gDNA Sequence sequence gDNA GDNA Cequence sequence general sequence sequence sequence sequence gDNA GDS reforence sequence sequence gDNA GDS reforence sequence sequence gDNA GDS reforence sequence sequence sequence sequence sequence sequence gDNA GDS reforence sequence	Table	2C: CYP	Table 2C: CYP3A5 polymorphisms	detected in samples of Chinese origin.	of Chir	nese origin.				
equence Reference sequence gDNA Seq reference sequence gDNA ID variant seq. element chzk 254T>G g20619T>G 052 TGGGCTGCAC 5 of PS2 chzk 582A>G g20291A>G 069 GCCCCACCTGG 5 of PS2 chzk 582A>G g6177A>G 069 CATCTTACCGG 5 of 1 chz 328T>C g1617T>C 070 CATCTTACCGG 5 of 1 chz 474G>A g1617T>C 070 CATCTTACCGG 5 of 1 chz 474G>A g30G>A 186 CATCTTACCGG 5 of 1 chz 474G>A g30G>A 187 CATCTTACCGGG 1 of 1 chz 474G>A g30G>A 146 TTTCACTATCGGGGG 1 inton 1 chz 441 441insCTAAAAAAT 399 CAGGGTATGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	Variant	Referenc	Variant po	sition on	Sed	Jence context	Genetic	Predicted	2	Variant allele
chzk 254T>G g20619T>G 051 TGGGCTGCGA 5' of PS2 chzk 582A>G g20291A>G 069 GCCCACCTCC 5' of PS2 chzl 229A>G g6177A>G 061 CTCACACTGGG 5' of PS2 chz 328T>C g1617T>C 062 CTCACACTGGG 5' of PS2 chzy 380T>C g1617T>C 069 CATATGCCTT Inton 1 chzy 474G>A g.230G>A 186 CTTATGCCTT Inton 1 chzy 474G>A g.230G>A 146 TTCAGTATT Inton 1 chzy 444insCTAAAAAAT g.6986G>A 146 TTCAGTATT Inton 4 chzu 444insCTAAAAAAT g.7424- 089 C-AG=G Inton 4 chzu 460CA g.12907G>A 188 ATACGGTATCT Inton 4 chzu 460CA g.13028G>A 146 TTCAGTATCA Inton 5 chzu 460CA g.13028G>A 192 AGGAGTATCA Inton 5 chz	<u> </u>	e e e		gDNA	Sed	reference seq.	element	effect		frequency (%)
chzk 582A-G g20291A-G 059 GCCCCACCTCC 5' of PS2 chzl 229A-G g6177A-G 061 CTCACACTGGG 5' of 1 chzl 328T-C g16177A-G 069 CATCTTACCC 5' of 1 chzy 380T-C g16177A-C 070 CTTATCCCTT Intron 1 chzy 474G-A g.230G-A 184 CCTTATCCCTT Intron 1 s chzy 474G-A g.230G-A 186 CTTATCCAGTATCT Intron 1 s chzy 441- 3.7424- 089 CT-AGTATCCAGTATCT Intron 3 s chzu 369G-A g.12907G-A 189 CT-AGTATCAGTATCT Intron 5 chzu 444insCTAAAAAT 039 CT-AGTATCATCT Intron 5 s chzu 444insCTAAAAAAT 039 CT-AGTATCATCT Intron 5 chzu 444insCTAAAAAAT 039 CT-AGTATCATCT Intron 5 chzu 440C-A g.12907G-A 189 AGTACGTTATC Intron 5 <td>ch-v-020</td> <td>chzk</td> <td>254T>G</td> <td>g20619T>G</td> <td>051</td> <td>TGGGCTTGCAA</td> <td>5' of PS2</td> <td></td> <td>42</td> <td>23.8</td>	ch-v-020	chzk	254T>G	g20619T>G	051	TGGGCTTGCAA	5' of PS2		42	23.8
chz 229A>G g6177A>G 061 CTCACACTGGG 5' of chyz 328T>C g1617T>C 062 G	ch-v-021	chzk	582A>G	g20291A>G	059	GCCCACCTCC	5' of PS2		45	26.7
chyz 328T>C g1617T>C 069 CATCTTACCCC 5 of chzy 380T>C g.136T>C 184 CCTTTTCCCTT Intron 1 chzy 474G>A g.230G>A 186 CTTATGCGTT Intron 1 chzy 474G>A g.6986G>A 146 TTTCAGTATTCT Intron 1 chzv 4441- g.7424- 089 C-AG-AGTATCT Intron 3 chzu 4441- 7427insCTAAAAAT 089 C-AG-AGTATCT Intron 4 chzu 440T- 7427insCTAAAAAT 089 C-AG-AGTATCT Intron 4 chzu 404T>C g.12907G>A 189 C-AG-AGTATGA Intron 5 chzu 404T>C g.12907G>A 189 C-AG-AGTATGA Intron 5 chzu 480G>A g.12952T>C 190 GGAGGTATGA Intron 5 chzu 480G>A g.17163G>T 192 AGGAGGTATGC Intron 9 chzu 6hzq g.27526C>T 106 CGAAACTACAT Intron 11 <	ch-v-026	chzl	229A>G	g6177A>G	061 062	CTCACACTGGG	5' of Exon 1		47	26.6
chzy 380T>C g.136T>C 184 CCTTTTCCCTT Intron 1 chzy 474G>A g.230G>A 186 CTTATGCAGAT Intron 1 chyu 206G>A g.6986G>A 146 TTTCAGTATCT Intron 1 chyu 206G>A g.6986G>A 146 TTTCAGTATCT Intron 3 chzu 441- g.7424- 030 CCTAAAAAT Intron 4 chzu 444insCTAAAAAT 7427insCTAAAAAT 030 CCTAAAAAAT Intron 4 chzu 404T>C g.12907G>A 189 AATACGGTCAT Exon 5 chzu 404T>C g.1295ZT>C 190 GGAGGTATCC Intron 5 chzu 480G>A g.13028G>A 190 GGAGGTATCC Intron 5 chzu 480G>A g.17163G>T 192 AGGAAGTATCC Intron 5 chzu g.13028G>A 193 CGAAACTACAT Intron 9 Intron 9 chz g.15 g.16 GGAAACTACT Intron 9 Intron 9 chz	ch-v-034	chyz	328T>C	g1617T>C	069 070	CATCTTACCCC	5' of Exon 1		47	21.3
chzy 474G>A g.230G>A 186 CTTATGCAGAT Intron 1 chyu 206G>A g.6986G>A 146 TTTCAGTATCT Intron 3 chzv 441- g.7424- 089 CAGTG Intron 4 chzu 359G>A G-AGTG Intron 4 chzu 404T>C g.12907G>A 189 ARTACGGTATGA chzu 404T>C g.12952T>C 190 GGAGGTATGA chzu 480G>A g.12952T>C 191 AGTCCGTTTCC chzu 480G>A g.13028G>A 192 AGTCCGTTTCC chzu 480G>A g.17163G>T 192 AGGAAGTATCC chzq 384G>T g.17163G>T 105 AGGAAGTATCC chzq 643C>T g.27526C>T 106 AGGAACTACAT Intron 11 chzn 351T>C g.31611T>C 107 AAGGAATTCTA Exon 13	ch-v-066	chzy	380T>C	g.136T>C	184 185	CCTTTTCCCTT	Intron 1		45	
chyu 206G>A g.6986G>A 146 TTTCAGTATCT Intron 3 chzv 441- g.7424- 089 CAGG Intron 4 chzu 444insCTAAAAAT 990 C-AGG Intron 4 chzu 359G>A g.12907G>A 188 AATACGGTCAT Exon 5 chzu 404T>C g.12952T>C 190 GGAGGTATGA Intron 5 chzu 480G>A g.13028G>A 192 AGTCCGTTTCC Intron 5 chzu 480G>A g.17163G>T 067 AGGAAGTATTC Intron 9 chzq 643C>T g.27526C>T 105 CGAAACTACAT Intron 11 chzn 642C>T 105 AGGAATTTCTA Exon 13 chzn 351T>C g.31611T>C 107 AAGGATTTCTA Exon 13	ch-v-067	chzy	474G>A	g.230G>A	186 187	CTTATGCAGAT	Intron 1		44	1.14
chzv 441- 444insCTAAAAAT g.7424- 7427insCTAAAAAAT 090 CC-AGG Intron 4 chzu 359G>A g.12907G>A 188 AATACGGTCAT Exon 5 chzu 404T>C g.1295ZT>C 190 GGAGGTATGAA Intron 5 chzu 480G>A g.13028G>A 192 AGTCCGTTTCC Intron 5 chzu 480G>A g.17163G>T 067 AGGAAGTATTC Intron 9 chzm 643C>T g.27526C>T 105 CGAAACTACAT Intron 11 chzn 351T>C g.31611T>C 107 AAGGATTTCTA Exon 13	ch-v-048	chyu	206G>A	g.6986G>A	1467	TTTCAGTATCT	Infron 3		. 47	26.6
chzu 359G>A g.12907G>A 188 AATACGGTCAT Exon 5 chzu 404T>C g.1295ZT>C 190 GGAGGTATGAA Intron 5 chzu 480G>A g.13028G>A 192 AGTCCGTTTCC Intron 5 chzu 480G>A g.17163G>T 067 AGGAAGTATTC Intron 9 chzm 643C>T g.27526C>T 105 CGAAACTACAT Intron 11 chzn 351T>C g.31611T>C 107 AAGGATTTCTA Exon 13	ch-v-018	chzv	441-	g.7424- 7427insCTAAAAAT	080 080	CAGG CCTAAAAAATG	Intron 4		47	2.1
chzu 404T>C g.12952T>C 190 GGAGGTATGAA Intron 5 chzu 480G>A g.13028G>A 192 AGTCCGTTTCC Intron 5 chzq 364G>T g.17163G>T 057 AGGAAGTATTC Intron 9 chzm 643C>T g.27526C>T 105 CGAAACTACAT Intron 11 chzn 351T>C g.31611T>C 107 AAGGATTTCTA Exon 13	ch-v-068	chzu	359G>A	g.12907G>A	188 189	AATACGGTCAT	Exon 5	R130Q	45	3.3
chzu 480G>A 9.13028G>A 192 AGTCCGTTTCC chzq 364G>T 9.17163G>T 057 AGGAAGTATTC chzm 643C>T 9.27526C>T 105 CGAAACTACAT chzm 351T>C G.31611T>C 107 AAGGATTTCTA	ch-v-047	chzu	404T>C	g,12952T>C	190 191	GGAGGTATGAA	Intron 5	splice defect	44	1.1
chzq 364G>T g.17163G>T 057 AGGAAGTATTC chzm 643C>T g.27526C>T 105 CGAAACTACAT chzm 351T>C g.31611T>C 107 AAGGATTTCTA	ch-v-069	chzu	480G>A	g.13028G>A	· 192 193	AGTCCGTTTCC	Intron 5		4	1.1
chzm 643C>T g.27526C>T 105 CGAAACTACAT chzn 351T>C g.31611T>C 107 AAGGATTTCTA	ch-v-011	chzq	364G>T	g.17163G>T	098 098	AGGAAGTATTC	Intron 9		40	21.3
chzn 351T>C g.31611T>C 107 AAGGATTTCTA	ch-v-014	chzm	643C>T	g.27526C>T	105 106	CGAAACTACAT	Intron 11		47	5.3
901	ch-v-015	chzn	351T>C	g.31611T>C	107	AAGGATTTCTA	Exon 13 (3' UTR)		47	26.6

Table	2D: CYP	Table 2D: CYP3A5 polymorphisms detected in samples of Japanese origin.	detected in samples	of Jap	ariese origin.				
Variant	Referenc	Variant po	Variant position on	Seq	Sequence context	Genetic	Predicted		Variant allele
Ō	ecuence	Reference sequence	gDÑA	Seq ID	reference seq.	element	effect	z	frequency (%
ch-v-020	chzk	254T>G	g20619T>G	051 052	regectrecaa	5' of PS2		42	29.8
ch-v-021	chzk	582A>G	920291A>G	020	GCCCACCTCC	5' of PS2		49	28.6
ch-v-026	chzl	229A>G	g6177A>G	061 062	CTCACACTGGG	5' of Exon 1		46	28.3
ch-v-028	chzh	601G>A	g3844G>A	990 066	TGTGTGTGGGA	5' of Exon 1		50	2.0
ch-v-034	chyz	328T>C	g1617T>C	069 070	CATC'FTACCCC	5' of Exon 1		50	26.0
ch-v-007	chzw	143C>T	g.5215C>T	083 084	GATAGCAGGCC	Intron 2		48	3.1
ch-v-048	chyu	206G>A	g.6986G>A	146 147	TTTCAGTATCT	Intron 3		50	29.0
ch-v-047	chzu	404T>C	g.12952T>C	190	GGAGGTATGAA	Intron 5	splice defect	50	1.0
ch-v-061	chzq	194C>G	g.16993C>G	194 195	TCTGCCAAAGA	Intron 8		49	1.0
ch-v-011	chzq	364G>T	g.17163G>T	097 098	AGGAAGTATTC	Intron 9		49	26.5
ch-v-017	chzm	248-249insT	g.27131-27132insT	111	CACCT-ACCTA	Exon 11	(D348.)	48	1.0
ch-v-014	chzm	643C>T	g.27526C>T	105 106	CGAAACTACAT	Intron 11		49	3.1
ch·v-045	chzn	291T>C	g.31551T>C	196 197.	ACCCATTGTTC	Exon 13	1488T	50	3.0
ch-v-015	chzn	351T>C	g.31611T>C	107 108	AAGGATTTCTA	Exon 13 (3' UTR)		50	31.0

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olymorphisms detected in samples of Korean origin.
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Table 2E: CYP3A5 polymorphisn
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Table
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Variant	Reference	riant Reference Variant position on	sition on	Sedu	Sequence context	Genetic	Predicted	2	Variant allele
Ω	ecuence	Reference sequence	gDNA	Sed	reference seq.	element	effect	2	frequency (%)
ch-v-020	chzk	254T>G	g20619T>G	051 052	TGGGCTTGCAA	5' of PS2		47	29.8
ch-v-065	chzk	563T>C	g20310T>C	198 199	GCTACTGGCTG	5' of PS2		47	1.1
ch-v-021	chzk	582A>G	g20291A>G	090	GCCCCACCTCC	5' of PS2		47	29.8
ch-v-040	chzl	206C>T	g6200C>T	200 201	GAAATCACCCG	5' of Exon 1		43	1.2
ch-v-026	chzi	229A>G	g6177A>G	061 062	CTCACACTGGG	5' of Exon 1		43	31.4
ch-v-034	chyz	328T>C	g1617T>C	069 070	CATCTTACCCC	5' of Exon 1		47	27.7
ch-v-007	chzw	143C>T	g.5215C>T	083 084	GATAGCAGGCC T	Intron 2		47	2.1
ch-v-048	chyu	206G>A	g.6986G>A	146 147	TTTCAGDATCT	Intron 3		47	29.8
ch-v-061	chzq	194C>G	g.16993C>G	194 195	TCTGCCAAAGA	Intron 8		47	2.1
ch-v-011	chzq	364G>T	g.17163G>T	097 098	AGGAAGTATTC T	Intron 9		47	27.7
ch-v-014	chzm	643C>T	g.27526C>T	105 106	CGAAACTACAT	Intron 11		47	2.1
ch-v-015	chzn	351T>C	g.31611T>C	107 - 108	AAGGATTTCTA	Exon 13 (3' UTR)		43	36.1

AC005020.2 as genomic reference sequences wherein the A of the ATG at position 166220 in AF280107.1 is +1. Sequence context: local alignment at the polymorphic site with the reference allele sequence given at the top and the variant sequence given below. Dots indicate nucleotide identity at the respective position. N: number of samples analysed. nomenclature is based on Antoarakis et al. (Antonarakis, Hum Mutat 11 (1998), 1-3) using the joined sequences AF280107.1 and Table 2A-E: Variants are listed according to their localisation along the gene, separately for each ethnic group. Polymorphism

Table 3: CYP3A5 genotypes and phenotypes.

	ch-v-021	ch-v-026	ch-v-015	Phenotype	Livers
Genotype 1	A/A	A/A	T/T	LE	155
Genotype 2	A/G	A/G	T/T	LE	9
Genotype 3	A/A	A/A	T/C	LE	4
Genotype 4	A/G	A/G	T/C	HE	17
Genotype 5	A/A	A/A	T/T	HE	1

All three variants were observed only in the heterozygous state. HE = high expressing livers, LE = low expressing livers. Numbers indicate LE and HE livers with each particular genotype. The increased CYP3A5 expression co-segregates with a distinct genotype.

Table 4: Expected drug metabolism by CYP3A5

Geno-		Enzyme		justment
type No.	Allelic combination	Activity	drug degradation	drug activation
1	high expressor allele/ high expressor allele	190 %	1.90	0.53
11, 111	low expressor allele/ high expressor allele	100 %	1.00	1.00
IV - VII	null allele/ high expressor allele	95 %	0.95	1.05
VIII, IX, X	low expressor allele/ low expressor allele	10 %	0.10	10
XI - XVIII	null allele/ low expressor allele	5 %	0.05	20
XIX - XXV	null allele/ null allele	0 %	< 0.05	> 20
Geno- type No.	Genotype (SeqiD) at Locus 1 - 2 - 3 - 4 - 5 - 6 - 7 - 8	Enzyme Activity	Dose Ad drug degradation	ustment drug activation
ì	060-062-079-081-087-111-103-108 / 060-062-079-081-087-111-103-108	190 %	1.90	0.53
II	0xx-06x-079-081-087-111-103-107 / 060-062-079-081-087-111-103-108	100 %	1.00	1.00 ,
Ш	059-061-079-081-087-111-103-10x /. 060-062-079-081-087-111-103-108	100 %	1.00	1.00
IV.	0xx-06x-080-082-08x-11x-10x-10x / 060-062-079-081-087-111-103-108	95 %	0.95	1.05
· V	0xx-06x-0xx-08x-088-11x-10x-10x / 060-062-079-081-087-111-103-108	95 %	0.95	1.05
VI	0xx-06x-0xx-08x-08x-112-10x-10x / 060-062-079-081-087-111-103-108	95 %	0.95	1.05
VII	0xx-06x-0xx-08x-08x-11x-104-10x / 060-062-079-081-087-111-103-108	95 %	0.95	1.05
VIII	060-062-079-081-087-111-103-107 / 059-061-079-081-087-111-103-108	10 %	0.10	10
IX	0xx-06x-079-081-087-111-103-107 / 0xx-06x-079-081-087-111-103-107	10 %	0.10	10
X	059-061-079-081-087-111-103-10x / 059-061-079-081-087-111-103-10x	10 %	0.10	10
ΧI	059-061-079-081-087-111-103-10x / 0xx-06x-080-082-08x-11x-10x-10x	5 %	0.05	20
XII	0xx-06x-079-081-087-111-103-107 / 0xx-06x-080-082-08x-11x-10x-10x	5 %	0.05	20
·XIII	059-061-079-081-087-111-103-10x / 0xx-06x-0xx-08x-088-11x-10x-10x	5 %	0.05	20
XIV	0xx-06x-079-081-087-111-103-107 / 0xx-06x-0xx-08x-088-11x-10x-10x	5 %	0.05	20
XV	059-061-079-081-087-111-103-10x / 0xx-06x-0xx-08x-08x-112-10x-10x	5 %	0.05	20
XVI	0xx-06x-079-081-087-111-103-107 /	5 %	0.05	20

				
	0xx-06x-0xx-08x-08x-112-10x-10x			
XVII	059-061-079-081-087-111-103-10x /	5 %	0.05	20
	0xx-06x-0xx-08x-08x-11x-104-10x			
XVIII	0xx-06x-079-081-087-111-103-107 /	5 %	0.05	20
	0xx-06x-0xx-08x-08x-11x-104-10x			
XIX	0xx-06x-080-082-08x-11x-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-088-11x-10x-10x			
XX	0xx-06x-080-082-08x-11x-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-08x-112-10x-10x			
XXI	0xx-06x-080-082-08x-11x-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-08x-11x-104-10x			
XXII	0xx-06x-080-082-08x-11x-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-080-082-08x-11x-10x-10x			
XXIII	0xx-06x-0xx-08x-088-11x-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-088-11x-10x-10x			
XXIV	0xx-06x-0xx-08x-08x-112-10x-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-08x-112-10x-10x			
XXV	0xx-06x-0xx-08x-08x-11x-104-10x /	0 %	< 0.05	> 20
	0xx-06x-0xx-08x-08x-11x-104-10x			

No.: running genotype number.

Genotype: Possible CYP3A5 genotypes that result from combinations of alleles. At the top of the table concise allele names have been used to indicate the principle. The lower table lists the alleles in greater detail, giving all combinations of variants at 8 loci in the two homologous chromosomes (loci 1 -8 refer to positions corresponding to positions -20291, -6177, 3705, 3709/3710, 7303, 27131/27132, 27289 and 31611, respectively, of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613), respectively. Each variant is defined by a 3-digit Seq ID as listed in Table 2A-E. A wildcard (x) in Seq IDs indicates that the phenotype is independent from the variant at this locus in this chromosome. The possible variants for each locus that can be substituted for x can be extracted from Table 2A-E. For example, 08x at locus 4 stands for Seq IDs 081 or 082, whereas 08x at locus 5 indicates either Seq ID 087 or 088.

Enzyme Activity: enzyme activity as calculated from protein concentration whereby the average protein concentration of genotype 059-061-079-081-111-107/060-062-079-081-111-108 was defined as 100 %.

Dose Adjustment: dose adjustment factors for drugs that are degraded/activated by CYP3A5 relative to the dosis required for genotype 059-061-079-081-111-107/060-062-079-081-111-108. Factors may need to be weighted according to the activity share of the CYP3A5 enzyme for drugs which are not exclusively metabolised by CYP3A5.

CLAIMS

- 1. A polynucleotide comprising a polynucleotide selected from the group consisting of:
 - (a) a polynucleotide having the nucleic acid sequence of SEQ ID NO: 54, 56, 58, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94, 96, 98, 100, 102, 106, 108, 110, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 128, 129, 130, 131, 133, 134, 135, 136, 137, 138, 139, 140, 142, 143, 149, 151, 153, 155, 157, 159, 161, 163, 165, 169, 171, 173, 175, 177, 179, 181, 183, 185, 187, 189, 193, 195, 197, 199, 201, 207, 208, 209, 210, 211, 212, 213, 214, 216, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 231, 232, 233, 235, or 236;
 - (b) a polynucleotide encoding a polypeptide having the amino acid sequence of SEQ ID NO: 127, 132, 141, 215, 229, or 234;
 - a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said (c) polynucleotide is having a nucleotide exchange, a nucleotide deletion of at least one nucleotide, or at least one additional nucleotide at a position corresponding to position -20643, -20555, -20359, -20367, -20329, -20323, -20310, -6200, -6177, -4336, -3990, -3868, -3844, -3557, -1617, -795, -86, -74, 136, 174 to 176, 230, 3705, 3709/3710, 5215, 5235, 5516, 7182, 7207, 7303, 7424/7427, 12907, 13028, 13077, 13173, 13226, 13376, 14720, 14836, 14903, 15788, 16079. 16993, 17163, 19069, 19165, 19208, 27050, 16931/16932. 27131/27132, 27526, 31499, 31551 or 31611;
 - a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said polynucleotide is having an A at a position corresponding to position -20555, -20329, -20323, -4336, -3868, -3844, -795, -86, 230, 5235, 5516, 7182, 7303, 12907, 13028, 13376, 19069 or 19165 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a T at a position corresponding to position -20367, -6200, -74, 3705, 5215, 7207, 14836, 17163, 19208 or 27526 of

the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a G at a position corresponding to position -6177, -3990, 13077, 14720, 14903, 16993 or 27050 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), a C at a position corresponding to position -20643, -20310, -3557, -1617, 136, 13173, 13226, 15788, 16079, 31499, 31551 or 31611 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614). nucleotide deletions at positions corresponding to positions 174 to 176 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613), an additional nucleotide at a position corresponding to position 3709/3710 or 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), three additional nucleotides at a position corresponding to position 16931/16932 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), or a deletion of two nucleotides and nine additional nucleotides inserted at a position corresponding to position 7424 to 7427 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613);

(e) a polynucleotide encoding a CYP3A5 polypeptide or fragment thereof, wherein said polypeptide comprises an amino acid substitution at a position corresponding to position 30, 100, 130, 149 or 488 of the

CYP3A5 polypeptide (Accession No: NP_000768.1), or at least one amino acid exchange or a stop codon at a position corresponding to position 30 to 34 or 346 to 348 of the CYP3A5 polypeptide (Accession No: NP_000768.1); and

- a polynucleotide encoding a CYP3A5 polypeptide or fragment thereof. (f) wherein said polypeptide comprises amino acid substitutions of HGLFK to YGTF, (with the period meaning termination) at a position corresponding to position 30 to 34 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid substitution of S to Y at a position corresponding to position 100 of the CYP3A5 polypeptide (Accession No: NP 000768.1), an amino acid substitution of R to Q at a position corresponding to position 130 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid substitution of I to T at a position corresponding to position 149 of the CYP3A5 polypeptide (Accession No: NP_000768.1), an amino acid substitutions of TYD to YL. (with the period meaning termination) at position corresponding to position 346 to 348 of the CYP3A5 polypeptide (Accession No: NP_000768.1), or an amino acid substitution of I to T at a position corresponding to position 488 of the CYP3A5 polypeptide (Accession No: NP_000768.1).
- 2. A polynucleotide of claim 1, wherein said polynucleotide is associated with cancer or diseases including cardiovascular diseases, diabetes and AIDS.
- 3. A polynucleotide of any one of claims 1 to 2 which is DNA or RNA.
- 4. A gene comprising the polynucleotide of any one of claims 1 to 2.
- 5. The gene of claim 4 wherein a nucleotide deletion, addition and/or substitution results in altered expression of the variant gene compared to the corresponding wild type gene.
- 6. A vector comprising a polynucleotide of any one of claims 1 to 3 or the gene of claim 4 or 5.

7. The vector of claim 6, wherein the polynucleotide is operatively linked to expression control sequences allowing expression in prokaryotic or eukaryotic cells or isolated fractions thereof.

- 8. A host cell genetically engineered with the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5 or the vector of claim 6 or 7.
- A method for producing a molecular variant CYP3A5 polypeptide or fragment thereof comprising
 - (a) culturing the host cell of claim 8; and
 - (b) recovering said protein or fragment from the culture.
- 10. A method for producing cells capable of expressing a molecular variant CYP3A5 polypeptide comprising genetically engineering cells with the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5 or the vector of claim 6 or 7.
- 11. A polypeptide or fragment thereof encoded by the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5 or obtainable by the method of claim 9 or from cells produced by the method of claim 10.
- 12. An antibody which binds specifically to the polypeptide of claim 11.
- 13. The antibody of claim 12 which specifically recognizes an epitope containing one or more amino acid substitution(s) resulting from a nucleotide exchange as defined in claim 1 or 5.
- 14. The antibody of claim 12 or 13 which is monoclonal or polyclonal.
- 15. A transgenic non-human animal comprising at least one polynucleotide of any one of claims 1 to 4, the gene of claim 5 or 6 or the vector of claim 7 or 8.

16. The transgenic non-human animal of claim 15 which is a mouse, a rat or a zebrafish.

- 17. A solid support comprising one or a plurality of the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5, the vector of claim 6 or 7, the polypeptide of claim 11, the antibody of claim 12 or 13 or the host cell of claim 8 in immobilized form.
- 18. The solid support of claim 17, wherein said solid support is a membrane, a glass-, polypropylene- or silicon-chip, are oligonucleotide conjugated beads or bead array, which is assembled on an optical filter substrate.
- 19. An in vitro method for identifying a polymorphism said method comprising the steps of:
 - (a) isolating a polynucleotide of any one claims 1 to 3 or the gene of claim 4 or 5 from a plurality of subgroups of individuals, wherein one subgroup has no prevalence for a CYP3A5 associated disease and at least one or more further subgroup(s) do have prevalence for a CYP3A5 associated disease; and
 - (b) identifying a polymorphism by comparing the nucleic acid sequence of said polynucleotide or said gene of said one subgroup having no prevalence for a CYP3A5 associated disease with said at least one or more further subgroup(s) having a prevalence CYP3A5 associated disease.
- 20. A method for identifying and obtaining a pro-drug or a drug capable of modulating the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:
 - (a) contacting the polypeptide of claim 11, the solid support of claim 17 or 18, a cell expressing a molecular variant gene comprising a polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5 or the vector of claim 6 or 7 in the presence of components capable of providing a detectable signal in response to drug activity with a compound to be screened for pro-drug or drug activity; and

(c) detecting the presence or absence of a signal or increase or decrease of a signal generated from the pro-drug or the drug activity, wherein the absence, presence, increase or decrease of the signal is indicative for a putative pro-drug or drug.

- 21. A method for identifying and obtaining an inhibitor of the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:
 - (d) contacting the protein of claim 11, the solid support of claim 17 or 18 or a cell expressing a molecular variant gene comprising a polynucleotide of any one of claims 1 to 3 or the gene of claim 4 or 5 or the vector of claim 6 or 7 in the presence of components capable of providing a detectable signal in response to drug activity with a compound to be screened for inhibiting activity; and
 - (e) detecting the presence or absence of a signal or increase or decrease of a signal generated from the inhibiting activity, wherein the absence or decrease of the signal is indicative for a putative inhibitor.
- 22. The method of claim 20 or 21, wherein said cell is a cell of claim 9, obtained by the method of claim 10 or can be obtained by the transgenic non-human animal of claim 15 or 16.
- 23. A method of identifying and obtaining a pro-drug or drug capable of modulating the activity of a molecular variant of a CYP3A5 polypeptide comprising the steps of:
 - (a) contacting the host cell of claim 8, the cell obtained by the method of claim 10, the polypeptide of claim 11 or the solid support of claim 17 or 18 with the first molecule known to be bound by a CYP3A5 polypeptide to form a first complex of said polypeptide and said first molecule;
 - (b) contacting said first complex with a compound to be screened, and
 - (c) measuring whether said compound displaces said first molecule from said first complex.

24. A method of identifying and obtaining an inhibitor capable of the activity of a molecular variant of a CYP3A5 polypeptide or its gene product comprising the steps of:

- (a) contacting the host cell of claim 8, the cell obtained by the method of claim 10, the protein of claim 11 or the solid support of claim 17 or 18 with the first molecule known to be bound by a CYP3A5 polypeptide to form a first complex of said protein and said first molecule;
- (b) contacting said first complex with a compound to be screened, and
- (c) measuring whether said compound displaces said first molecule from said first complex.
- 25. The method of claim 23 or 24, wherein said measuring step comprises measuring the formation of a second complex of said protein and said compound.
- 26. The method of any one of claim 23 to 25, wherein said measuring step comprises measuring the amount of said first molecule that is not bound to said protein.
- 27. The method of any one of claims 23 to 26, wherein said first molecule is labeled.
- 28. A method for the production of a pharmaceutical composition comprising the steps of the method of any one of claims 20 to 27; and the further step of formulating the compound identified and obtained or a derivative thereof in a pharmaceutically acceptable form.
- 29. A method of diagnosing a disorder related to the presence of a molecular variant of a CYP3A5 gene or susceptibility to such a disorder comprising determining the presence of a polynucleotide of any one of claims 1 to 3 or the gene of claim 4 or 5 in a sample from a subject.
- 30. The method of claim 29 further comprising determining the presence of a polypeptide of claim 11 or the antibody of any one of claims 12 to 14.

31. A method of diagnosing a disorder related to the presence of a molecular variant of a CYP3A5 gene or susceptibility to such a disorder comprising determining the presence of a polypeptide of claim 11 or the antibody of any one of claims 12 to 14 in a sample from a subject.

- 32. The method of any one of claims 29 to 31, wherein said disorder is cancer or diseases including cardiovascular diseases, diabetes and AIDS.
- 33. The method of any one of claims 29 to 32 comprising PCR, ligase chain reaction, restriction digestion, direct sequencing, nucleic acid amplification techniques, hybridization techniques mass spectroscopy or immunoassays.
- 34. A method of detection of the polynucleotide of any one of claims 1 to 3 or the gene of claim 4 or 5 in a sample comprising the steps of
 - (a) contacting the solid support of claim 17 or 18 with the sample under conditions allowing interaction of the polynucleotide of claim 1 to 3 or the gene of claim 4 or 5 with the immobilized targets on a solid support and;
 - (b) determining the binding of said polynucleotide or said gene to said immobilized targets on a solid support.
- 35. An in vitro method for diagnosing a disease comprising the steps of the method of claim 34, wherein binding of said polynucleotide or gene to said immobilized targets on said solid support is indicative for the presence or the absence of said disease or a prevalence for said disease.
- 36. A diagnostic composition comprising the polynucleotide of any one of claims 1 to 4, the gene of claim 3 or 4, the vector of claim 6 or 7, the polypeptide of claim 11 or the antibody of claim 12 or 13.
- 37. A pharmaceutical composition comprising the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5, the vector of claim 6 or 7, the polypeptide of claim 11 or the antibody of claim 12 or 13.

38. Use of the polynucleotide of any one of claims 1 to 3, a polynucleotide comprising SEQ ID No: 104, a polynucleotide encoding a polypeptide comprising SEQ ID No: 145, the gene of claim 4 or 5, the vector of claim 6 or 7, the polypeptide of claim 11, a polypeptide comprising SEQ ID No: 145 or the antibody of claim 12 or 13 for the preparation of a diagnostic composition for diagnosing a disease.

- 39. Use of the polynucleotide of any one of claims 1 to 3, a polynucleotide comprising SEQ ID No: 104, a polynucleotide encoding a polypeptide comprising SEQ ID No: 145, the gene of claim 4 or 5, the vector of claim 6 or 7, the polypeptide of claim 11, a polypeptide comprising SEQ ID No: 145 or the antibody of claim 12 or 13 for the preparation of a pharmaceutical composition for treating a disease.
- 40. Use of a polynucleotide selected from the group consisting of:
 - a polynucleotide having the nucleic acid sequence of SEQ ID NO: 82,
 88, 104 or 112;
 - (b) a polynucleotide encoding a polypeptide having the amino acid sequence of SEQ ID No: 127, 132, 141 or 145;
 - a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said polynucleotide is having at least one additional nucleotide at a position corresponding to position 3709/3710 or 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614) or a nucleotide exchange at a position corresponding to position 7303 or 27289 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614);
 - (d) a polynucleotide capable of hybridizing to a CYP3A5 gene, wherein said polynucleotide is having an additional G nucleotide at a position

corresponding to position 3709/3710 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), an additional T nucleotide at a position corresponding to position 27131/27132 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614), or an A at a position corresponding to position 7303 or 27289 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614);

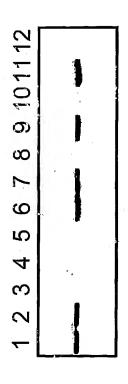
for the preparation of a diagnostic composition for diagnosing a disease in a subject having a genome comprising a variant allele of the CYP3A5 gene, wherein said allele is having an A at a position corresponding to position 6986 of the CYP3A5 gene (Accession No. AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No. AC005020.2, wherein position 27341 has been numbered +8614).

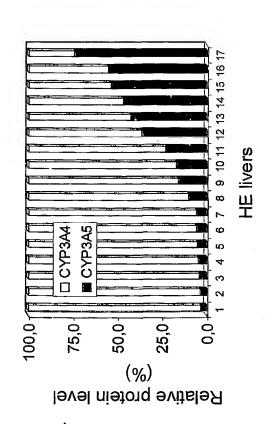
41. Use of a polynucleotide comprising a polynucleotide having an A at a position corresponding to position 14690 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614) for the preparation of a diagnostic composition for diagnosing a disease in a subject having a genome comprising a variant allele of the CYP3A5 gene, wherein said allele is having an A at a position corresponding to position 6986 of the CYP3A5 gene (Accession No: AF280107.1, wherein position 166220 has been numbered +1 and position 174832 has been numbered +8613, and Accession No: AC005020.2, wherein position 27341 has been numbered +8614).

42. The use of claim 40 or 41, wherein said subject is an African American.

- 43. The use of any one of claims 38 to 41, wherein said disease is cancer or diseases including cardiovascular diseases, diabetes and AIDS.
- 44. A diagnostic kit for detection of a single nucleotide polymorphism comprising the polynucleotide of any one of claims 1 to 3, the gene of claim 4 or 5, the vector of claim 6 or 7, the polypeptide of claim 11, the antibody of claim 12 or 13, the host cell of claim 8, the transgenic non-human animal of claim 15 or 16 or the solid support of claim 17 or 18.







2/21 Figure 2

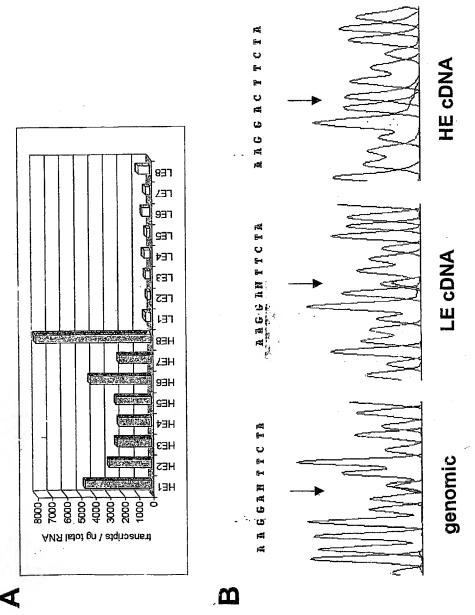


Figure 3

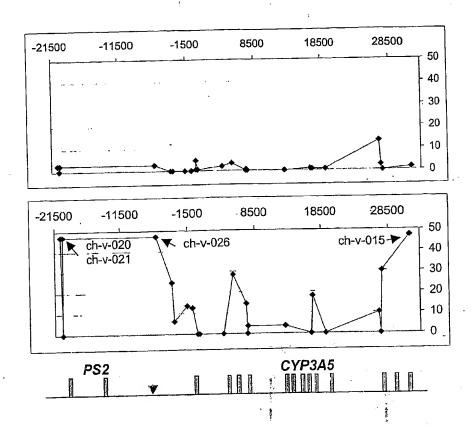


Figure 4

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Figure 4 continued

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-/214

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gaaacataga gaacggttgc tactggcaga agcataagat ctttgtacaa tattgctggc 720
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ttgagetete ccagagtetg tgtagagtgt tgtgcattat gtagtataaa ggaggtgace 540
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<213> Homo sapiens
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agcaaacagc agcactcagc taaaaggaag actcacagaa cacagttgaa gaaggaaagt 240
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aagtagetga agtgttggac getacaaacg catagaagtt attattatet tatgcagate 480
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caatgaaaac atggcagcgg caga
<210> 227
<211> 624
<212> DNA
<213> Homo sapiens
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aagtagctga agtgttggac gctacaaacg catagaagtt attattatct tatacagatc 480
tatgaatgaa taaataagca tttctcccat ccaccttcta attttggtga ctaggagggt 540
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Ile Ile Ala Gln Tyr Gly Asp Val Leu Val Arg Asn
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gaggetagge atctaggate tecattgage atcttgaata tggettgeea taatateata 180
tacagtcaat aaatatttgt taaataagga tgcctcttca atatattttg tgcaaccatg 240
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Gln Asn Phe Ser Phe Lys Pro Cys Lys Glu Thr Gln Ile Pro Leu Lys
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ataaaqcagg ctgtgaagag gggattccca tgctcgtgtg cctgataaca caactatcac 720
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1 .		\$. ⁵	i i i
47/47			

4/47